PRIONS

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by

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ABSTRACT

Prions are unprecedented infectious pathogens that cause a group of invariably fatal neurodegenerative diseases by an entirely novel mechanism. Prion diseases may present as genetic, infectious, or sporadic disorders, all of which involve modification of the prion protein (PrP). Bovine spongiform encephalopathy (BSE), scrapie of sheep, and Creutzfeldt-Jakob disease (CJD) of humans are among the most notable prion diseases. Prions are transmissible particles that are devoid of nucleic acid and seem to be composed exclusively of a modified protein (PrP^{Sc}). The normal, cellular PrP (PrP^{C}) is converted into PrP^{Sc} through a posttranslational process during which it acquires a high β-sheet content. The species of a particular prion is encoded by the sequence of the chromosomal PrP gene of the mammals in which it last replicated. In contrast to pathogens carrying a nucleic acid genome, prions appear to encipher strain-specific properties in the tertiary structure of PrP^{Sc}. Transgenetic studies argue that PrP^{Sc} acts as a template upon which PrP^{C} is refolded into a nascent PrP^{Sc} molecule through a process facilitated by another protein. Miniprions generated in transgenic mice expressing PrP, in which nearly half of the residues were deleted, exhibit unique biological properties and should facilitate structural studies of PrP^{Sc}. While knowledge about prions has profound implications for studies of the structural plasticity of proteins, investigations of prion diseases suggest that new strategies for the prevention and treatment of these disorders may also find application in the more common degenerative diseases.

The tortuous path of the scientific investigation that led to an understanding of familial Creutzfeldt-Jakob disease (CJD) chronicles a remarkable scientific odyssey. By 1930, the high incidence of familial (f) CJD in some families was known [1,2]. Almost 60 years were to pass before the significance of this finding could be appreciated [3-5]. CJD remained a curious, rare neurodegenerative disease of unknown etiology throughout this period of three score years [6]. Only with transmission of disease to apes following inoculation of brain extracts prepared from patients, who died of CJD, did the story begin to unravel [7].

Once CJD was shown to be an infectious disease, relatively little attention was paid to the familial form of the disease since most cases were not found in

families. It is interesting to speculate how the course of scientific investigation might have proceeded had transmission studies not been performed until after the molecular genetic lesion had been identified. Had that sequence of events transpired, then the prion concept, which readily explains how a single disease can have a genetic or infectious etiology, might have been greeted with much less skepticism [8].

Epidemiologic studies designed to identify the source of the CJD infection were unable to identify any predisposing risk factors although some geographic clusters were found [9–12]. Libyan Jews living in Israel developed CJD about 30 times more frequently than other Israelis [13]. This finding prompted some investigators to propose that the Libyan Jews had contracted CJD by eating lightly cooked brain from scrapie-infected sheep when they lived in Tripoli prior to emigration. Subsequently, the Libyan Jewish patients were all found to carry a mutation at codon 200 in their prion protein (PrP) gene [14–16].

My own interest in the subject began with a patient dying of CJD in the fall of 1972. At that time, I was beginning a residency in Neurology and was most impressed by a disease process that could kill my patient in two months by destroying her brain while her body remained unaffected by this process. No febrile response, no leucocytosis or pleocytosis, no humoral immune response, and yet I was told that she was infected with a "slow virus."

Slow viruses. The term "slow virus" had been coined by Bjorn Sigurdsson in 1954 while he was working in Iceland on scrapie and visna of sheep [17]. Five years later, William Hadlow had suggested that kuru, a disease of New Guinea highlanders, was similar to scrapie and thus, it, too, was caused by a slow virus [18]. Seven more years were to pass before the transmissibility of kuru was established by passaging the disease to chimpanzees inoculated intracerebrally [19]. Just as Hadlow had made the intellectual leap between scrapie and kuru, Igor Klatzo made a similar connection between kuru and CJD [20]. In both instances, these neuropathologists were struck by the similarities in light microscopic pathology of the CNS that kuru exhibited with scrapie or CJD. In 1968, the transmission of CJD to chimpanzees after intracerebral inoculation was reported [7].

In scrapie, kuru, CJD, and all of the other disorders now referred to as prion diseases (Table 1), spongiform degeneration and astrocytic gliosis is found upon microscopic examination of the CNS (Figure 1) [21]. The degree of spongiform degeneration is quite variable while the extent of reactive gliosis correlates with the degree of neuron loss [22].

Prions–a brief overview. Before proceeding with a detailed discussion of our current understanding of prions causing scrapie and CJD, I provide a brief overview of prion biology. Prions are unprecedented infectious pathogens that cause a group of invariably fatal neurodegenerative diseases mediated by an entirely novel mechanism. Prion diseases may present as genetic, infectious, or sporadic disorders, all of which involve modification of the prion protein (PrP), a constituent of normal mammalian cells [23]. CJD generally presents as progressive dementia while scrapie of sheep and bovine spongiform encephalopathy (BSE) are generally manifest as ataxic illnesses (Table 1) [24].

Table 1. The prion diseases

|	Disease	Host	Mechanism of Pathogenesis
A.	Kuru	Fore people	Infection through ritualistic cannibalism
	Iatrogenic CJD*	Humans	Infection from prion-contaminated HGH, dura mater grafts, etc.
	Variant CJD	Humans	Infection from bovine prions?
	Familial CJD	Humans	Germline mutations in PrP gene
	GSS	Humans	Germline mutations in PrP gene
	FFI	Humans	Germline mutation in PrP gene (D178N, M129)
	Sporadic CJD	Humans	Somatic mutation or spontaneous conversion of PrP^C into PrP^{Sc}?
	FSI	Humans	Somatic mutation or spontaneous conversion of PrP^C into PrP^{Sc}?
B.	Scrapie	Sheep	Infection in genetically susceptible sheep
	BSE	Cattle	Infection with prion-contaminated MBM
	TME	Mink	Infection with prions from sheep or cattle
	CWD	Mule deer, elk	Unknown
	FSE	Cats	Infection with prion-contaminated bovine tissues or MBM
	exotic ungulate encephalopathy	Greater kudu, nyala, onyx	Infection with prion-contaminated MBM

*Abbreviations defined in Footnote a.
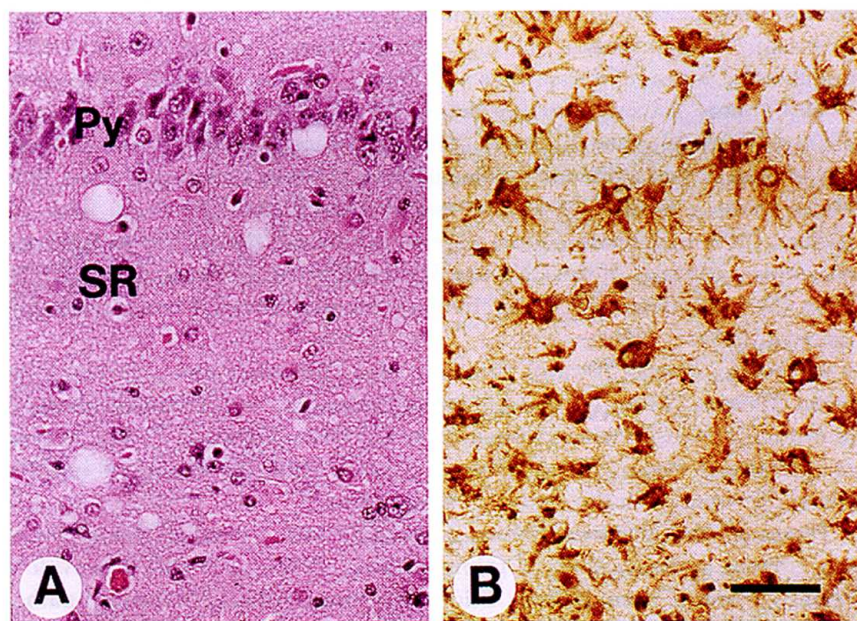
Figure 1. Neuropathologic changes in Swiss mice following inoculation with RML scrapie prions. (A) Hematoxylin and eosin stain of a serial section of the hippocampus shows spongiform degeneration of the neuropil, with vacuoles 10–30 µm in diameter. Brain tissue was immersion fixed in 10% buffered formalin solution after sacrificing the animals and embedded in paraffin. (B) GFAP immunohistochemistry of a serial section of the hippocampus shows numerous reactive astrocytes. Bar in B = 50 µm and also applies to A. Photomicrographs prepared by Stephen J. DeArmond.

Prions are devoid of nucleic acid and seem to be composed exclusively of a modified isoform of PrP designated PrP^{Sc}.[a] The normal, cellular PrP, denoted PrP^C, is converted into PrP^{Sc} through a process whereby a portion of its α-helical and coil structure is refolded into β-sheet [25]. This structural transition is accompanied by profound changes in the physicochemical properties of the PrP. The amino acid sequence of PrP^{Sc} corresponds to that encoded by the PrP gene of the mammalian host in which it last replicated. In contrast to pathogens with a nucleic acid genome that encode strain-specific properties in genes, prions encipher these properties in the tertiary structure of PrP^{Sc} [26–28]. Transgenetic studies argue that PrP^{Sc} acts as a template upon which PrP^C is refolded into a nascent PrP^{Sc} molecule through a process facilitated by another protein.

More than 20 mutations of the PrP gene are now known to cause the inherited human prion diseases and significant genetic linkage has been established for five of these mutations [4, 16, 29–31]. The prion concept readily explains how a disease can be manifest as a heritable as well as an infectious illness.

Resistance of scrapie agent to radiation. My fascination with CJD quickly shifted to scrapie once I learned of the remarkable radiobiological data that Tikvah Alper and her colleagues had collected on the scrapie agent [32–34]. The scrapie agent had been found to be extremely resistant to inactivation by UV and ionizing radiation as was later shown for the CJD agent [35]. It seemed to me that the most intriguing question was the chemical nature of the scrapie agent; Alper's data had evoked a torrent of hypotheses concerning its composition. Suggestions as to the nature of the scrapie agent ranged from small DNA viruses to membrane fragments to polysaccharides to proteins, the last of which eventually proved to be correct [36–42].

Scrapie of sheep and goats possesses a history no less fascinating than that of CJD. The resistance of the scrapie agent to inactivation by formalin and heat treatments [43], which were commonly used to produce vaccines against viral illnesses, suggested that the scrapie agent might be different from viruses, but it came at a time before the structure of viruses was understood. Later, this resistance was dismissed as an interesting observation but of little importance since some viruses can survive such treatments; indeed, this was not an unreasonable viewpoint. More than two decades were to pass before reports of the extreme resistance of the scrapie agent to inactivation by radiation again trumpeted the novelty of this infectious pathogen. Interestingly, British scientists had argued for many years about whether natural scrapie was a genetic or an infectious disease [44–46]. Scrapie, like kuru and CJD, produced

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[a] Prions are defined as proteinaceous infectious particles that lack nucleic acid. PrP^C is the cellular prion protein; PrP^{Sc} is the pathologic isoform. NH$_2$-terminal truncation during limited proteolysis of PrP^{Sc} produces PrP 27-30. BSE, bovine spongiform encephalopathy; CJD, Creutzfeldt-Jakob disease; sCJD, sporadic CJD; fCJD, familial CJD; iCJD, iatrogenic CJD; vCJD, (new) variant CJD; CWD, chronic wasting disease; FFI, fatal familial insomnia; FSE, feline spongiform encephalopathy; FSI, fatal sporadic insomnia; GSS, Gerstmann-Sträussler-Scheinker disease; HGH, human growth hormone; MBM, meat and bone meal; SHa, Syrian hamster; Tg, transgenic; TME, transmissible mink encephalopathy; wt, wild-type.

death of the host without any sign of an immune response to a "foreign infectious agent."

My initial studies focused on the sedimentation properties of scrapie infectivity in mouse spleens and brains. From these studies, I concluded that hydrophobic interactions were responsible for the non-ideal physical behavior of the scrapie particle [47, 48]. Indeed, the scrapie agent presented a biochemical nightmare: infectivity was spread from one end to the other of a sucrose gradient and from the void volume to fractions eluting at 5–10 times the included volume of chromatographic columns. Such results demanded new approaches and better assays [49].

Bioassays. As the number of hypotheses about the molecular nature of the scrapie agent began to exceed the number of laboratories working on this problem, the need for new experimental approaches became evident. Much of the available data on the properties of the scrapie agent had been gathered on brain homogenates prepared from mice with clinical signs of scrapie. These mice had been inoculated 4–5 months earlier with the scrapie agent which originated in sheep but had been passaged multiple times in mice. Once an experiment was completed on these homogenates, an additional 12 months was required using an endpoint titration in mice [50]. Typically, 60 mice were required to determine the titer of a single sample. This slow, tedious, and expensive system discouraged systematic investigation.

Although the transmission of scrapie to mice had ushered in a new era of research, the 1.5–2 year intervals between designing experiments and obtaining results discouraged sequential studies. Infrequently, the results of one set of experiments were used as a foundation for the next and so on. Moreover, the large number of mice needed to measure the infectivity in a single sample prevented studies where many experiments were performed in parallel. These problems encouraged publication of inconclusive experimental results.

In 1972, when I became fascinated by the enigmatic nature of the scrapie agent, I thought that the most direct path to determining the molecular structure of the scrapie agent was purification. Fortunately, I did not appreciate the magnitude of that task although I had considerable experience and training in the purification of enzymes [51]. While many studies had been performed to probe the physicochemical nature of the scrapie agent using the mouse endpoint titration system, few systematic investigations had been performed on the fundamental characteristics of the infectious scrapie particle [42]. In fact, 12 years after introduction of the mouse bioassay, there were few data on the sedimentation behavior of the scrapie particle. Since differential centrifugation is frequently a useful initial step in the purification of many macromolecules, some knowledge of the sedimentation properties of the scrapie agent under defined conditions seemed mandatory. To perform such studies, Swiss mice were inoculated intracerebrally with the Chandler isolate of scrapie prions and the mice sacrificed about 30 and 150 days later when the titers in their spleens and brains, respectively, were expected to be at maximal levels. The two tissues were homogenized, extracted with detergent, and

centrifuged for increasing times and speeds [47, 52]. The disappearance of scrapie infectivity was measured in supernatant fractions by endpoint titration which required one year to score.

Incubation time assays in hamsters. In view of these daunting logistical problems, the identification of an inoculum that produced scrapie in the golden Syrian hamster in ~70 days after intracerebral inoculation proved to be an important advance [53, 54] once an incubation time assay was developed [55, 56]. In earlier studies, Syrian hamsters (SHa) had been inoculated with prions, but serial passage with short incubation times was not reported [57]. Development of the incubation time bioassay reduced the time required to measure prions in samples with high titers by a factor of five: only 70 days were required instead of the 360 days previously needed. Equally important, 4 animals could be used in place of the 60 that were required for endpoint titrations, making possible a large number of parallel experiments. With this bioassay, research on the nature of the scrapie agent was accelerated nearly 100-fold and the hamster with high prion titers in its brain became the experimental animal of choice for biochemical studies.

The incubation time assay enabled development of effective purification schemes for enriching fractions for scrapie infectivity. It provided a means to assess quantitatively those fractions that were enriched for infectivity and those that were not. Such studies led rather rapidly to the development of a protocol for separating scrapie infectivity from most proteins and nucleic acids. With a ~100-fold purification of infectivity relative to protein, >98% of the proteins and polynucleotides were eliminated permitting more reliable probing of the constituents of these enriched fractions.

The prion concept. As reproducible data began to accumulate indicating that scrapie infectivity could be reduced by procedures that hydrolyze or modify proteins but was resistant to procedures that alter nucleic acids, a family of hypotheses about the molecular architecture of the scrapie agent began to emerge [58]. These data established, for the first time, that a particular macromolecule was required for infectivity and that this macromolecule was a protein. The experimental findings extended earlier observations on resistance of scrapie infectivity to UV irradiation at 250 nm [33] in that the four different procedures used to probe for a nucleic acid are based on physical principles that are independent of UV radiation damage.

Once the requirement of protein for infectivity was established, I thought that it was appropriate to give the infectious pathogen of scrapie a provisional name that would distinguish it from both viruses and viroids. After some contemplation, I suggested the term "prion" derived from **pro**teinaceous and **in**fectious [58]. At that time, I defined prions as proteinaceous infectious particles that resist inactivation by procedures that modify nucleic acids. I never imagined the irate reaction of some scientists to the word prion – it was truly remarkable!

Current definitions. Perhaps, the best current working definition of a prion is a proteinaceous infectious particle that lacks nucleic acid [28]. Because a wealth of data supports the contention that scrapie prions are composed entirely of

a protein that adopts an abnormal conformation, it is not unreasonable to define prions as infectious proteins [25, 27, 59, 60]. But I hasten to add that we still cannot eliminate a small ligand bound to PrP^{Sc} as an essential component of the infectious prion particle. Learning how to renature PrP^{Sc} accompanied by restoration of prion infectivity or to generate de novo prion infectivity using a synthetic polypeptide should help address this as yet unresolved issue [61]. From a broader perspective, prions are elements that impart and propagate conformational variability.

Although PrP^{Sc} is the only *known* component of the infectious prion particles, these unique pathogens share several phenotypic traits with other infectious entities such as viruses. Because some features of the diseases caused by prions and viruses are similar, some scientists have difficulty accepting the existence of prions despite a wealth of scientific data supporting this concept [62-67].

Families of hypotheses. Once the requirement for a protein was established, it was possible to revisit the long list of hypothetical structures that had been proposed for the scrapie agent and to eliminate carbohydrates, lipids, and nucleic acids as the infective elements within a scrapie agent devoid of protein [58]. No longer could structures such as a viroid-like nucleic acid, a replicating polysaccharide, or a small polynucleotide surrounded by a carbohydrate be entertained as reasonable candidates to explain the puzzling properties of the scrapie agent [58, 68].

The family of hypotheses that remained after identifying a protein component was still large and required a continued consideration of all possibilities in which a protein was a critical element [49]. The prion concept evolved from a family of hypotheses in which an infectious protein was only one of several possibilities. With the accumulation of experimental data on the molecular properties of the prion, it became possible to discard an increasing number of hypothetical structures. In prion research, as in many other areas of scientific investigation, a single hypothesis is all too often championed at the expense of a reasoned approach which requires entertaining a series of complex arguments until one or more can be discarded on the basis of experimental data [69].

Genes and DNA. In some respects, the early development of the prion concept mirrors the story of DNA [70-72]. Prior to the acceptance of DNA as the genetic material of life [73, 74], many scientists asserted that the DNA preparations must be contaminated with protein which is the true genetic material [75]. The prejudices of these scientists were similar in some ways to those of investigators who have disputed the prion concept. But the scientists who attacked the hypothesis that genes are composed of DNA had no well proven alternative; they had only a set of feelings derived from poorly substantiated data sets that genes are made of protein. In contrast, those who attacked the hypothesis that the prion is composed only of protein had more than 30 years of cumulative evidence showing that genetic information in all organisms on our planet is encoded in DNA and that biological diversity resides in DNA. Studies of viruses and eventually viroids extended this concept to these small

infectious pathogens [76] and showed that genes could also be composed of RNA [77, 78].

Discovery of the Prion Protein. The discovery of the prion protein transformed research on scrapie and related diseases [79, 80]. It provided a molecular marker that was subsequently shown to be specific for these illnesses as well as the major, and very likely the only, constituent of the infectious prion.

PrP 27–30 was discovered by enriching fractions from SHa brain for scrapie infectivity [79, 80]. This protein is the protease-resistant core of PrP^{Sc} and has an apparent molecular weight (M_r) of 27–30 kD [81, 82]. Although resistance to limited proteolysis proved to be a convenient tool for many but not all studies, use of proteases to enrich fractions for scrapie infectivity created a problem when the NH_2-terminal sequence of PrP 27–30 was determined [81]. The ragged NH_2-terminus of PrP 27–30 yielded three sets of signals in almost every cycle of the Edman degradation. Only once these signals were properly interpreted and placed in correct register could a unique sequence be assigned for the NH_2-terminus of PrP 27–30. The determination of amino acid sequence of the NH_2-terminus of PrP 27–30 made subsequent molecular cloning studies of the PrP gene possible [83, 84].

The finding that PrP mRNA levels were similar in normal uninfected and scrapie-infected tissues caused some investigators to argue that PrP 27–30 was not related to the infectious prion particle [83]. An alternate interpretation prompted a search for a prion protein in uninfected animals which was found to be protease sensitive and soluble in non-denaturing detergents, unlike PrP 27–30. This isoform was designated PrP^C (Figure 2) [84, 85]. Deduced amino acid sequences from PrP cDNA as well as immunoblotting studies revealed that PrP 27–30 was NH_2-terminally truncated and was derived from a larger molecule, designated PrP^{Sc}, that was unique to infected animals [81, 82, 84–86].

With the discovery of PrP 27–30 and the production of antiserum [87], brains from humans and animals with putative prion diseases were examined for the presence of this protein. In each case, PrP 27–30 was found and it was absent in other neurodegenerative disorders such as Alzheimer's disease, Parkinson's disease, and amyotrophic lateral sclerosis [88–91]. The extreme specificity of PrP^{Sc} for prion disease is an important feature of the protein and is consistent with the postulated role of PrP^{Sc} in both the transmission and pathogenesis of these illnesses (Table 2) [92].

The accumulation of PrP^{Sc} contrasts markedly with that of glial fibrillary acid protein (GFAP) in prion disease. In scrapie, GFAP mRNA and protein levels rise as the disease progresses [93] but the accumulation of GFAP is neither specific nor necessary for either the transmission or pathogenesis of disease. Mice deficient for GFAP show no alteration in their incubation times [94, 95].

Except for PrP^{Sc}, no macromolecule has been found in tissues of patients dying of the prion diseases that is specific for these encephalopathies. In searches for a scrapie-specific nucleic acid, cDNAs have been identified that are complementary to mRNAs encoding other proteins with increased expression in prion disease [96–98]. Yet, none of the proteins has been found to be specific for prion disease.
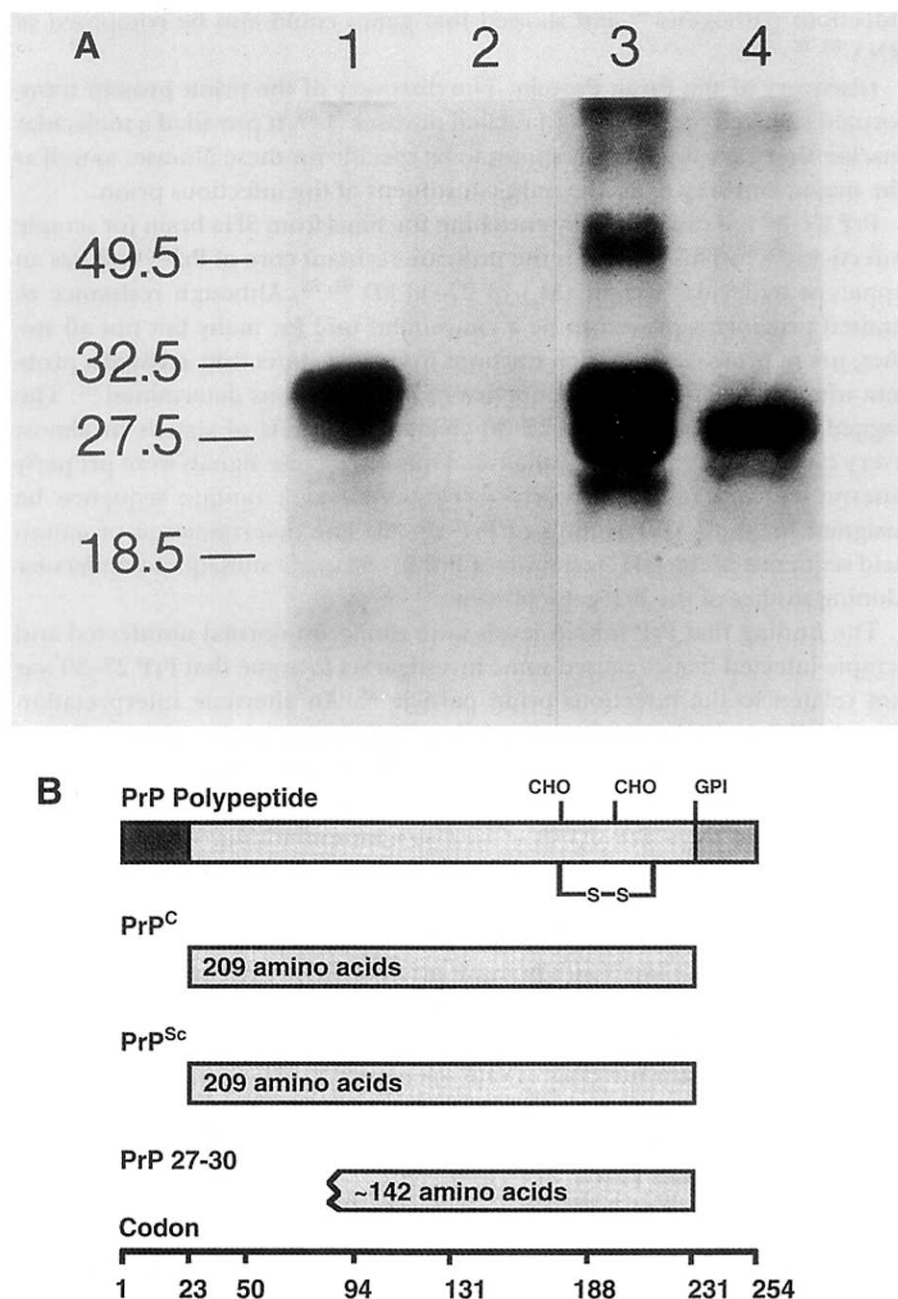
Figure 2. Prion protein isoforms. **(A)** Western immunoblot of brain homogenates from uninfect-ed (lanes 1 and 2) and prion-infected (lanes 3 and 4) Syrian hamsters. Samples in lanes 2 and 4 were digested with 50 µg/ml of proteinase K for 30 min at 37°C. PrP^C in lanes 2 and 4 was com-pletely hydrolyzed under these conditions, whereas approximately 67 amino acids were digested from the NH_2-terminus of PrP^{Sc} to generate PrP 27–30. After polyacrylamide gel electrophoresis (PAGE) and electrotransfer, the blot was developed with anti-PrP R073 polyclonal rabbit antise-rum. Molecular size markers are in kilodaltons (kD). **(B)** Bar diagram of SHaPrP which consists of 254 amino acids. After processing of the NH_2- and COOH- termini, both PrP^C and PrP^{Sc} con-sist of 209 residues. After limited proteolysis, the NH_2-terminus of PrP^{Sc} is truncated to form PrP 27–30, which is composed of approximately 142 amino acids.

Table 2. Arguments for prions being composed largely, if not entirely, of PrP^{Sc} molecules and devoid of nucleic acid.

1. PrP^{Sc} and scrapie infectivity copurify using biochemical and immunologic procedures.
2. The unusual properties of PrP^{Sc} mimic those of prions. Many different procedures that modify or hydrolyze PrP^{Sc} inactivate prions.
3. Levels of PrP^{Sc} are directly proportional to prion titers. Non-denatured PrP^{Sc} has not been separated from scrapie infectivity.
4. No evidence for a virus-like particle or a nucleic acid genome.
5. Accumulation of PrP^{Sc} invariably associated with the pathology of prion diseases including PrP amyloid plaques that are pathognomonic.
6. PrP gene mutations are genetically linked to inherited prion disease and cause formation of PrP^{Sc}.
7. Overexpression of PrP^{C} increases the rate of PrP^{Sc} formation, which shortens the incubation time. Knock out of the PrP gene eliminates the substrate necessary for PrP^{Sc} formation and prevents both prion disease and prion replication.
8. Species variations in the PrP sequence are responsible, at least in part, for the species barrier which is found when prions are passaged from one host to another.
9. PrP^{Sc} preferentially binds to homologous PrP^{C} resulting in formation of nascent PrP^{Sc} and prion infectivity.
10. Chimeric and partially deleted PrP genes change susceptibility to prions from different species and support production of artificial prions with novel properties that are not found in nature.
11. Prion diversity is enciphered within the conformation of PrP^{Sc}. Strains can be generated by passage through hosts with different PrP genes. Prion strains are maintained by PrP^{C}/PrP^{Sc} interactions.
12. Human prions from fCJD(E200K) and FFI patients impart different properties to chimeric MHu2M PrP in transgenic mice, which provides a mechanism for strain propagation.

Attempts to falsify the prion hypothesis. Numerous attempts to disprove the prion hypothesis over the past 15 years have failed. Such studies have tried unsuccessfully to separate scrapie infectivity from protein and more specifically from PrP^{Sc}. No preparations of purified prions containing less than one PrP^{Sc} molecule per ID$_{50}$ unit have been reported [99] and no replication of prions in PrP deficient (Prnp$^{0/0}$) mice was found [100-104].

Copurification of PrP 27–30 and scrapie infectivity demands that the physicochemical properties as well as antigenicity of these two entities be similar [105] (Table 2). The results of a wide array of inactivation experiments demonstrated the similarities in the properties of PrP 27–30 and scrapie infectivity [61, 106-109]. To explain these findings in terms of the virus hypothesis, it is necessary to postulate either a virus that has a coat protein which is highly homologous with PrP or a virus that binds tightly to PrP^{Sc}. In either case, the PrP-like coat protein or the PrP^{Sc}/virus complex must display properties indistinguishable from PrP^{Sc} alone. With each species that the putative virus invades, it must incorporate a new PrP sequence during replication.

Search for a scrapie-specific nucleic acid. The inability to inactivate preparations highly enriched for scrapie infectivity by procedures that modify nucleic acids militates against the existence of a scrapie-specific nucleic acid [58, 110, 111]. To explain the findings in terms of a virus, one must argue that PrP^{Sc} or an as yet undetected PrP-like protein of viral origin protects the viral genome from

inactivation. The notion that the putative scrapie virus encodes a PrP-like protein was refuted by nucleic acid hybridization studies using a PrP cDNA probe. Less than 0.002 nucleic acid molecules encoding PrP per ID_{50} unit were found in purified preparations of SHa prions [84]. To circumvent this finding, it could be hypothesized that the genetic code used by the PrP gene differs so greatly from that found in the cell that a PrP cDNA probe failed to detect it in highly purified preparations.

If prions contained a genome with a unique genetic code, then it is likely that this genome would encode some specialized proteins required for replication as well as some unique tRNAs. But both UV and ionizing radiation inactivation studies as well as physical studies have eliminated the possibility of a large nucleic acid hiding within purified preparations of prions [110–112]. Only oligonucleotides of fewer than 50 bases were found at a concentration of one molecule per ID_{50} unit in prion preparations highly enriched for scrapie infectivity [113, 114]. These small nucleic acids were of variable length and are thought to be degradation byproducts generated during purification of prions. Failure to find a bona fide genome was attributed to the unusual properties of the putative viral nucleic acid or technical incompetence on the part of the investigators who were unable to find it [63, 115].

PrP Amyloid. In preparations highly enriched for scrapie infectivity and containing only PrP 27–30 by silver staining of gels after SDS-PAGE, numerous rod-shaped particles were seen by electron microscopy after negative staining (Figure 3) [107]. Each of the rods was slightly different, in contrast to viruses which exhibit extremely uniform structures [116]. These irregular rods
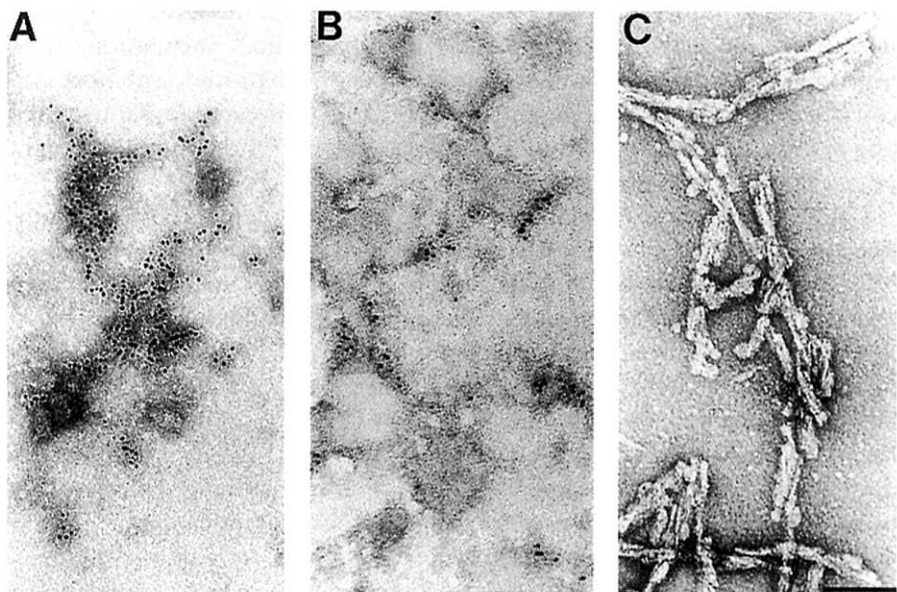

Figure 3. Electron micrographs of negatively stained and immunogold-labeled prion proteins: **(A)** PrP^C and **(B)** PrP^{Sc}. Neither PrP^C nor PrP^{Sc} form recognizable, ordered polymers. **(C)** Prion rods composed of PrP 27–30 were negatively stained. The prion rods are indistinguishable from many purified amyloids. Bar = 100 nm.

composed largely, if not entirely, of PrP 27–30, were indistinguishable morphologically from many other purified amyloids [117]. Studies of the prion rods with Congo red dye demonstrated that the rods also fulfilled the tinctorial criteria for amyloid [107] and immunostaining later showed that PrP is a major component of amyloid plaques in some animals and humans with prion disease [118–120]. Subsequently, it was recognized that the prion rods were not required for scrapie infectivity [121]; furthermore, the rods were shown to be an artifact of purification during which limited proteolysis of PrP^{Sc} generated PrP 27–30 that polymerized spontaneously in the presence of detergent (Figure 3) [122].

The idea that scrapie prions were composed of an amyloidogenic protein was truly heretical when it was introduced [107]. Since the prevailing view at the time was that scrapie is caused by an atypical virus, many argued that amyloid proteins are mammalian polypeptides and not viral proteins!

Some investigators have argued that the prion rods are synonymous with scrapie-associated fibrils [123–125] even though morphologic and tinctorial features of these fibrils clearly differentiated them from amyloid and as such from the prion rods [126, 127]. The scrapie-associated fibrils were identified by their unique ultrastructure in which two or four subfilaments were helically wound around each other [126] and were proposed to represent the first example of a filamentous animal virus [128]. After the argument for a filamentous animal virus causing scrapie faded, it was hypothesized that a virus induces the formation of PrP amyloid in order to explain the accumulation of PrP^{Sc} in prion diseases [129]. Besides the lack of evidence for a virus of any shape, no compelling data has been offered in support of the idea that prion diseases are caused by a filamentous bacterium called a spiroplasma [130].

Search for the ubiquitous "scrapie virus." When PrP gene mutations were discovered to cause familial prion diseases [4], it was postulated that PrP^{C} is a receptor for the ubiquitous scrapie virus that binds more tightly to mutant than to wt PrP^{C} [131]. A similar hypothesis was proposed to explain why the length of the scrapie incubation time was found to be inversely proportional to the level of PrP expression in Tg mice and why Prnp$^{0/0}$ mice are resistant to scrapie [132]. The higher the level of PrP expression, the faster the spread of the putative virus which results in shorter incubation times; conversely, mice deficient for PrP lack the receptor required for spread of the virus [63]. The inability to find virus-like particles in purified preparations of PrP^{Sc} was attributed to these particles being hidden [115] even though tobacco mosaic viruses could be detected when one virion was added per ID$_{50}$ unit of scrapie prions [121].

Recent studies on the transmission of mutant prions from FFI and fCJD(E200K) to Tg(MHu2M) mice which results in the formation of two different PrP^{Sc} molecules [27] has forced a corollary to the ubiquitous virus postulate. To accommodate this result, at least two different viruses must reside worldwide, each of which binds to a different mutant HuPrP^{C} and each of which induces a different MHu2M PrP^{Sc} conformer when transferred to Tg mice. Even more difficult to imagine is how one ubiquitous virus might acquire different mutant PrP^{Sc} molecules corresponding to FFI or fCJD(E200K)

and then induce different MHu2M PrP^{Sc} conformers upon transmission to Tg mice.

Artificial prions. To explain the production of artificial prions from chimeric or mutant PrP transgenes in terms of a virus [133–135], mutated PrP^{Sc} molecules must be incorporated into the virus. In the case of mice expressing chimeric PrP transgenes, artificial prions are produced with host ranges not previously found in nature. Similarly, deleting specific regions of PrP resulted in the formation of "miniprions" with a unique host range and neuropathology as described below. The production of artificial prions that were generated by modifying the PrP gene sequence and exhibit unique biological properties is another compelling argument against the proposition that scrapie and CJD are caused by viruses.

Skepticism once well-justified. While the skepticism about prions was once well-justified and formed the basis for a vigorous scientific debate, the wealth of available data now renders such arguments moot. In summary, no single hypothesis involving a virus can explain the findings summarized above (Table 2); instead, a series of ad hoc hypotheses, virtually all of which can be refuted by experimental data, must be constructed to accommodate a steadily enlarging body of data.

It is notable that the search for an infectious pathogen with a nucleic acid genome as the cause of scrapie and CJD has done little to advance our understanding of these diseases. Instead, studies of PrP have created a wealth of data that now explain almost every aspect of these fascinating disorders. While no single experiment can refute the existence of the "scrapie virus", all of the data taken together from numerous experimental studies present an impressive edifice which argues that the 50 year quest for a virus has failed because it does not exist!

Prions defy rules of protein structure. Once cDNA probes for PrP became available, the PrP gene was found to be constitutively expressed in adult, uninfected brain [83, 84]. This finding eliminated the possibility that PrP^{Sc} stimulated production of more of itself by initiating transcription of the PrP gene as proposed nearly two decades earlier [37]. Determination of the structure of the PrP gene eliminated a second possible mechanism that might explain the appearance of PrP^{Sc} in brains already synthesizing PrP^{C}. Since the entire protein coding region was contained within a single exon, there was no possibility for the two PrP isoforms to be the products of alternatively spliced mRNAs [82]. Next, a posttranslational chemical modification that distinguishes PrP^{Sc} from PrP^{C} was considered but none was found in an exhaustive study [59] and we considered it likely that PrP^{C} and PrP^{Sc} differed only in their conformation, a hypothesis also proposed earlier [37]. However, this idea was no less heretical than that of an infectious protein.

For more than 25 years, it had been widely accepted that the amino acid sequence specifies one biologically active conformation of a protein [136]. Yet, in scrapie we were faced with the possibility that one primary structure for PrP might adopt at least two different conformations in order to explain the existence of both PrP^{C} and PrP^{Sc}. When the secondary structures of the PrP iso-

forms were compared by optical spectroscopy, they were found to be markedly different [25]. Fourier transform infrared (FTIR) and circular dichroism (CD) studies showed that PrP^C contains about 40% α-helix and little β-sheet while PrP^{Sc} is composed of about 30% α-helix and 45% β-sheet [25, 137]. Nevertheless, these two proteins have the same amino acid sequence!

Prior to comparative studies on the structures of PrP^C and PrP^{Sc}, we found by metabolic labeling studies that the acquisition of PrP^{Sc} protease-resistance is a posttranslational process [138]. In our quest for a chemical difference that would distinguish PrP^{Sc} from PrP^C, we found ethanolamine in hydrolysates of PrP 27–30 which signalled the possibility that PrP might contain a glycosyl-phosphatidyl inositol (GPI) anchor [139]. Both PrP isoforms were found to carry GPI anchors and PrP^C was found on the surface of cells where it could be released by cleavage of the anchor. Subsequent studies showed that PrP^{Sc} formation occurred after PrP^C reaches the cell surface [140, 141] and is localized to caveolae-like domains [142–145].

Modeling PrP structures. Molecular modeling studies predicted that PrP^C is a four-helix bundle protein containing four regions of secondary structure denoted H1, H2, H3, and H4 (Figure 4) [146, 147]. Subsequent NMR studies of a synthetic PrP peptide containing residues 90–145 provided good evidence for H1 [148]. This peptide contains the residues 113–128 that are most highly conserved among all species studied (Figure 4A) [147, 149, 150] and correspond to a transmembrane region of PrP which was delineated in cell-free translation studies [151, 152]. Recent studies show that a transmembrane form of PrP accumulates in GSS caused by the A117V mutation and in Tg mice overexpressing either mutant or wtPrP [153]. The paradoxical lack of evidence for an α-helix in this region from NMR studies of recombinant PrP in aqueous buffers [154–156] could be explained if the recombinant PrPs correspond to the secreted form of PrP that was also identified in cell-free translation studies. This contention is supported by studies with recombinant antibody fragments (Fabs) showing the GPI-anchored PrP^C on the surface of cells exhibits an immunoreactivity similar to that of recombinant PrP that was prepared with an α-helical conformation [157, 158]. GPI-anchored PrP^C is synthesized within the secretory pathway and transported to surface of the cell [139, 159].

Optical spectroscopic measurements of PrP^C provided the necessary background for more detailed structural studies [25]. Unable to produce crystals of PrP, we and others utilized NMR to determine the structure of an α-helical form of a recombinant PrP. The NMR structure of a COOH-terminal fragment of MoPrP consisting of 111 residues showed three helices, two of which corresponded to H3 and H4 in the PrP^C model, and two small β-strands each consisting of three residues [154]. How the structure of this MoPrP(121–231) fragment differs from PrP^C is of interest since this fragment is lethal when expressed in Tg mice [160]. Subsequently, structural studies were performed on a longer fragment of PrP containing residues 90–231 and corresponding to SHaPrP 27–30 [155, 161, 162]. Expression of PrP(90–231) in Tg mice did not produce spontaneous disease [163, 164]. More recently, NMR structures of recombinant full-length PrP have been reported [156, 165].
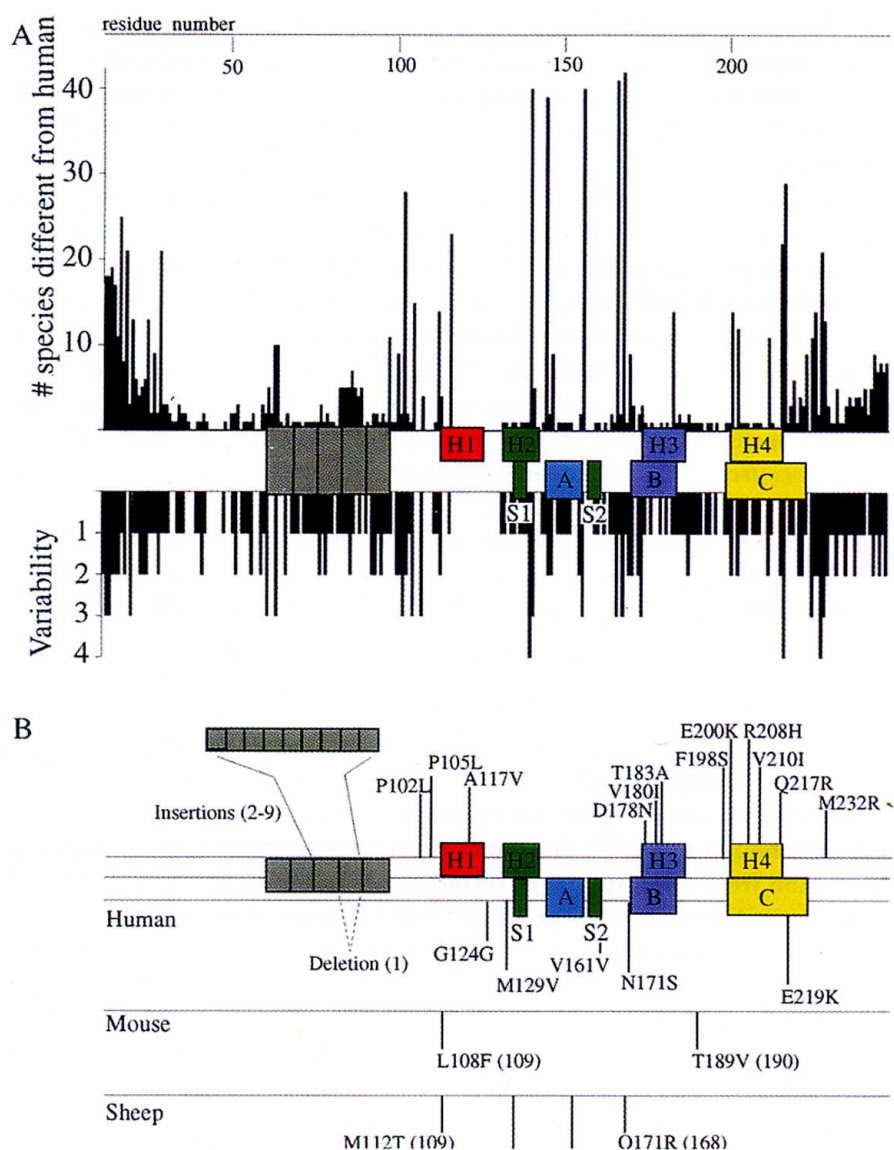
Figure 4. Species variations and mutations of the prion protein gene. (A) Species variations. The x-axis represents the human PrP sequence, with the five octarepeats and H1–H4 regions of putative secondary structure shown as well as the three α-helices A, B, and C and the two β-strands S1 and S2 as determined by NMR. The precise residues corresponding to each region of secondary structure are given in Figure 5. Vertical bars above the axis indicate the number of species that differ from the human sequence at each position. Below the axis, the length of the bars indicates the number of alternative amino acids at each position in the alignment. (B) Mutations causing inherited human prion disease and polymorphisms in human, mouse, and sheep. Above the line of the human sequence are mutations that cause prion disease. Below the lines are polymorphisms, some but not all of which are known to influence the onset as well as the phenotype of disease. Data were compiled by Paul Bamborough and Fred E. Cohen.

Models of PrP^Sc suggested that formation of the disease-causing isoform involves refolding of residues within the region between residues 90 and 140 into β-sheets [166]; the single disulfide bond joining COOH-terminal helices

would remain intact since the disulfide is required for PrP^{Sc} formation (Figure 5E) [167, 168]. The high β-sheet content of PrP^{Sc} was predicted from the ability of PrP 27–30 to polymerize into amyloid fibrils [107]. Subsequent optical

Figure 5A
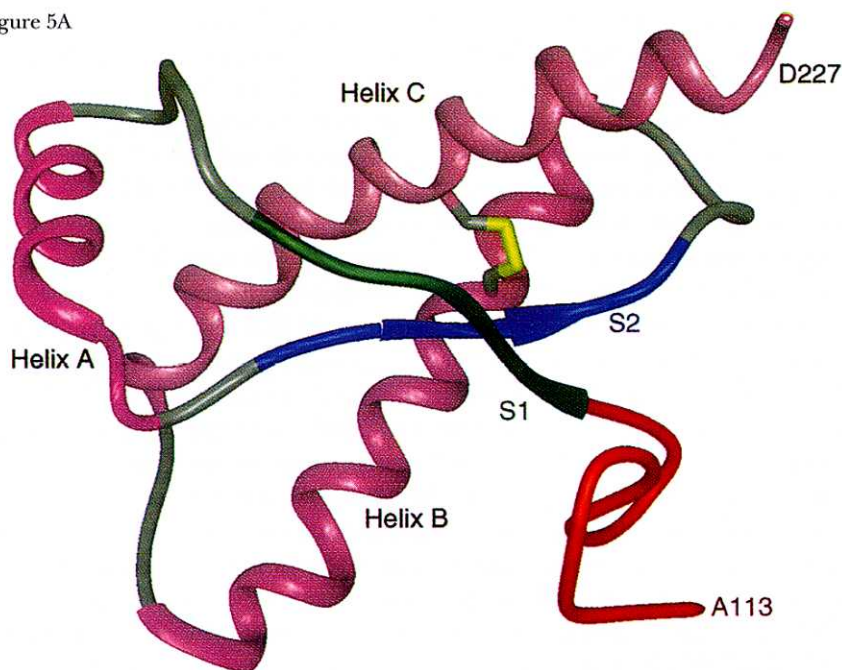

Figure 5. Structures of prion proteins. (A) NMR structure of Syrian hamster (SHa) recombinant (r) PrP(90–231). Presumably, the structure of the α-helical form of rPrP(90–231) resembles that of PrP^C. rPrP(90–231) is viewed from the interface where PrP^{Sc} is thought to bind to PrP^C. The color scheme is: α-helices A (residues 144–157), B (172–193), and C (200–227) in pink; disulfide between Cys179 and Cys214 in yellow; conserved hydrophobic region composed of residues 113–126 in red; loops in gray; residues 129–134 in green encompassing strand S1 and residues 159–165 in blue encompassing strand S2; the arrows span residues 129–131 and 161–163, as these show a closer resemblance to β–sheet [155]. (B) NMR structure of rPrP(90–231) is viewed from the interface where protein X is thought to bind to PrP^C. Protein X appears to bind to the side chains of residues that form a discontinuous epitope: some amino acids are in the loop composed of residues 165–171 and at the end of helix B (Gln168 and Gln172 with a low density van der Waals rendering) while others are on the surface of helix C (Thr215 and Gln219 with a high density van der Waals rendering) [178]. (C) PrP residues governing the transmission of prions [298]. NMR structure of recombinant SHaPrP region 121–231 [155] shown with the putative epitope formed by residues 184, 186, 203, and 205 highlighted in red. Residue numbers correspond to SHaPrP. Additional residues (138, 139, 143, 145, 148, and 155) that might participate in controlling the transmission of prions across species are depicted in green. Residues 168, 172, 215, and 219 that form the epitope for the binding of protein X are shown in blue. The three helices (A, B, and C) are highlighted in pink. (D) Schematic diagram showing the flexibility of the polypeptide chain for PrP(29–231) [156]. The structure of the portion of the protein representing residues 90–231 was taken from the coordinates of PrP(90–231) [155]. The remainder of the sequence was hand-built for illustration purposes only. The color scale corresponds to the heteronuclear {[1H]-[15N]} NOE data: red for the lowest (most negative) values, where the polypeptide is most flexible, to blue for the highest (most positive) values in the most structured and rigid regions of the protein. (E) Plausible model for the tertiary structure of human PrP^{Sc} [166]. Color scheme is: S1 β–strands are 108–113 and 116–122 in red; S2 β-strands are 128–135 and 138–144 in green; a-helices H3 (residues 178–191) and H4 (residues 202–218) in gray; loop (residues 142–177) in yellow. Four residues implicated in the species barrier are shown in ball-and-stick form (Asn 108, Met 112, Met 129, Ala 133).

spectroscopy confirmed the presence of β-sheet in both PrP[Sc] and PrP 27–30 [25, 169–171]. Deletion of each of the regions of putative secondary structure in PrP, except for the NH$_2$-terminal 66 amino acids (residues 23–88) [163, 172] and a 36 amino acid region (Mo residues 141 -176) prevented formation of PrP[Sc] as measured in scrapie–infected cultured neuroblastoma cells [168]. With α-PrP Fabs selected from phage display libraries [157] and two monoclonal antibodies (mAb) derived from hybridomas [173–175], the major conformational change that occurs during conversion of PrP[C] into PrP[Sc] has been localized largely, but not entirely, to a region bounded by residues 90 and 112 [158]. Similar conclusions were drawn from studies with an α-PrP IgM mAb [176]. While these results indicate that PrP[Sc] formation involves primarily a conformational change in the domain comprised of residues 90–112, mutations causing inherited prion diseases have been found throughout the protein (Figure 4B). Interestingly, all of the known point mutations in PrP with biological significance occur either within or adjacent to regions of putative secondary structure in PrP and as such, appear to destabilize the structure of PrP [147, 148, 154].

NMR structure of recombinant PrP. The NMR structure of recombinant SHaPrP(90–231) derived from *E. coli* was determined after the protein was purified and refolded (Figure 5A). Residues 90–112 are not shown since marked conformational heterogeneity was found in this region while residues 113–126 constitute the conserved hydrophobic region that also displays some structural plasticity [162]. Although some features of the structure of rPrP(90–231) are similar to those reported earlier for the smaller recombinant MoPrP(121–231) fragment [154, 177], substantial differences were found.
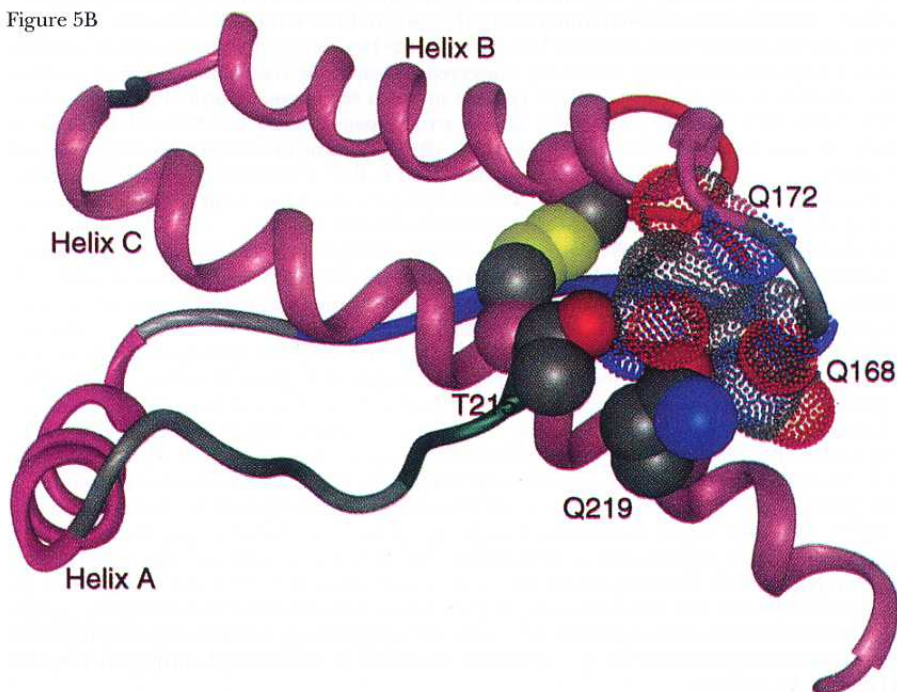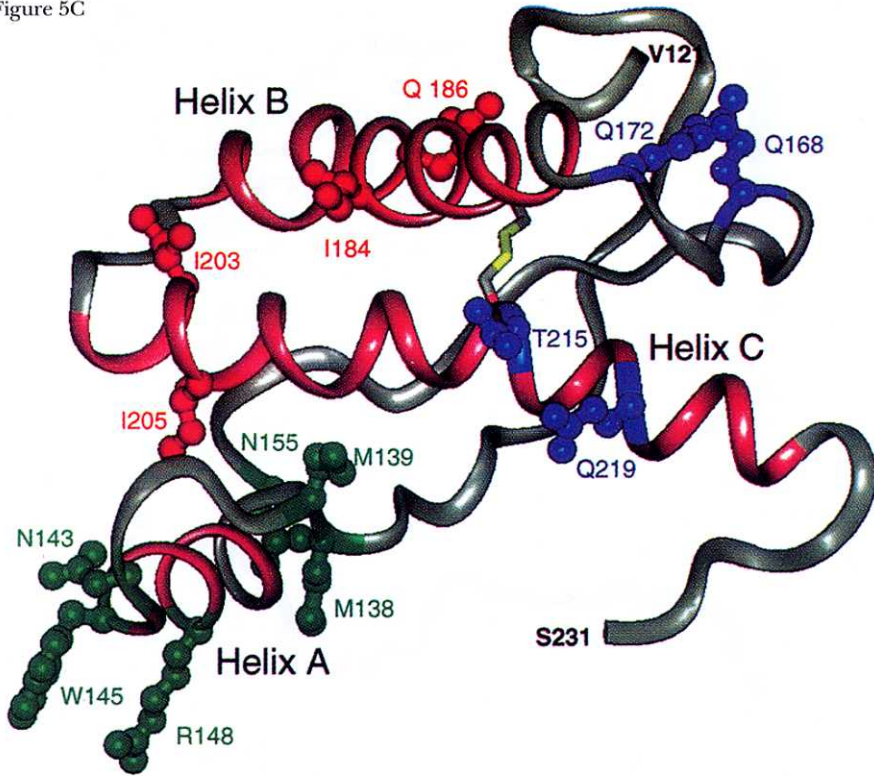
Figure 5B

Figure 5C

For example, the loop at the NH_2-terminus of helix B is well defined in rPrP(90–231) but is disordered in MoPrP(121–231); in addition, helix C is composed of residues 200–227 in rPrP(90–231) but extends only from 200–217 in MoPrP(121–231). The loop and the COOH-terminal portion of helix C are particularly important since they form the site to which protein X binds as described below (Figure 5B) [178]. Whether the differences between the two recombinant PrP fragments are due to (i) their different lengths, (ii) species specific differences in sequences, or (iii) the conditions used for solving the structures remains to be determined.

Studies of chimeric SHa/Mo and Hu/Mo PrP transgenes identified a domain comprised of residues 95 –170 where PrP^C binds to PrP^{Sc} [133, 179]. When chimeric Bo/Mo PrP transgenes failed to render mice sensitive to BSE prions, we examined the differences among the sequences in the chimeric and parent PrP genes [298]. The findings identified a second domain in PrP composed of residues 180–205 which seems to modulate the interaction between PrP^C and PrP^{Sc} (Figure 5C).

Recent NMR studies of full length MoPrP(23–231) and SHaPrP(29–231) have shown that the NH_2-termini are highly flexible and lack identifiable secondary structure under the experimental conditions employed (Figure 5D) [156, 165]. Studies of SHaPrP(29–231) indicate transient interactions between the COOH-terminal end of helix B and the highly flexible, NH_2-terminal ran-

Figure 5D
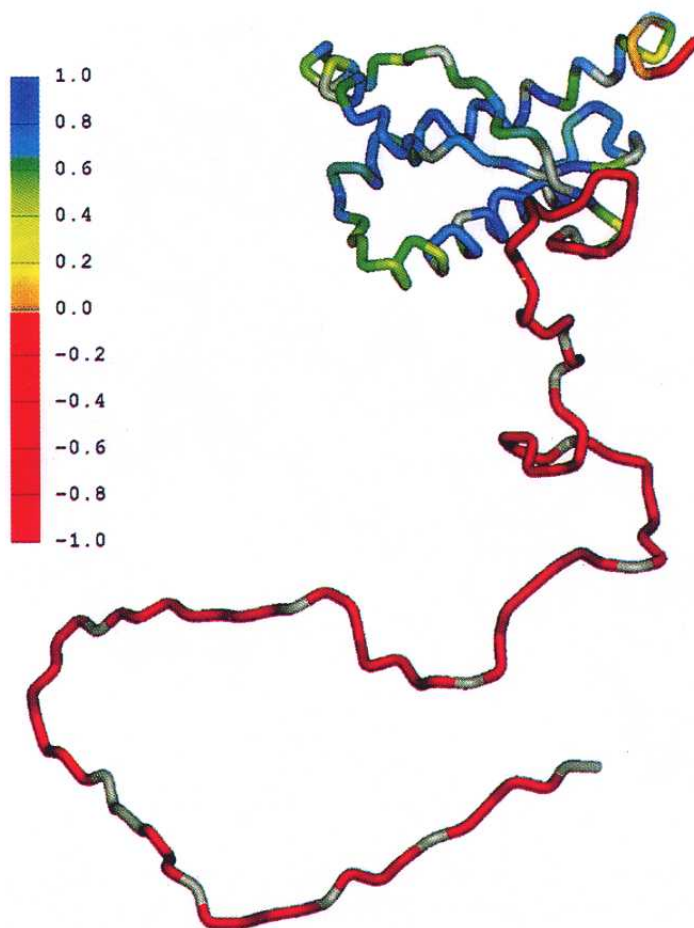
dom-coil containing the octareapeats (residues 29–125) [156]; such interactions were not reported for MoPrP(23–231) [165].

PrP appears to bind copper. The highly flexible NH_2-terminus of recombinant PrP may be more structured in the presence of copper. Each SHaPrP(29–231) molecule was found to bind two atoms of Cu^{2+}; other divalent cations did not bind to PrP [180]. Earlier studies with synthetic peptides corresponding to the octarepeat sequence demonstrated the binding of Cu^{2+} ions [181, 182] and optical spectroscopy showed that Cu^{2+} induced an α-helix formation in these peptides [183]. More recently, PrP-deficient ($Prnp^{0/0}$) mice were found to have lower levels of Zn/Cu superoxide dismutase (SOD) activity than controls [184]; SOD activity has been shown to mirror the state of copper metabolism [185]. Measurements of membrane extracts from brains of $Prnp^{0/0}$ mice showed low levels of Cu whereas Fe and Zn were unchanged suggesting PrP^C might function as a Cu^{2+} binding protein [186].

Disturbances in Cu^{2+} homeostasis leading to dysfunction of the CNS are well documented in humans and animals but are not known to be due to abnormalities in PrP metabolism: Menkes disease is manifest at birth and is due to a mutation of the MNK gene on the X chromosome while Wilson's disease

Figure 5E
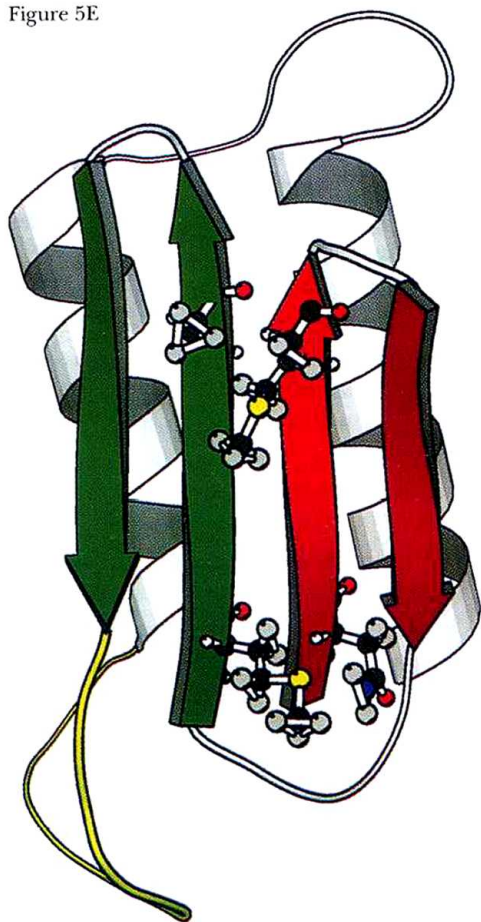
appears in childhood and is due to a mutation of the WD gene on chromosome 13 [187–190]. Both the MNK and WD genes encode copper transporting ATPases. While both Menkes and Wilson's diseases are recessive disorders, only Wilson's disease can be treated with copper chelating reagents. Interestingly, cuprizone, a Cu^{2+} chelating reagent, has been used in mice to induce neuropathological changes similar to those found in the prion diseases [191, 192].

PrP gene structure and expression. The entire open reading frame (ORF) of all known mammalian and avian PrP genes resides within a single exon [4, 82, 193, 194]. The mouse (Mo), sheep, cattle, and rat PrP genes contain three exons with the ORFs in exon 3 [195–199] which is analogous to exon 2 of the Syrian hamster (SHa) gene [82]. The two exons of the Syrian hamster (SHa) PrP gene are separated by a 10-kb intron: exon 1 encodes a portion of the 5' untranslated leader sequence while exon 2 encodes the ORF and 3' untranslated region [82]. Recently, a low abundance SHaPrP mRNA containing an additional small exon in the 5' untranslated region was discovered that is encoded by the SHaPrP gene [200]. Comparative sequencing of sheep and human (Hu) cosmid clones containing PrP genes revealed an additional putative, small untranslated 5' exon in the HuPrP gene [201]. The promoters of both the

SHa and Mo PrP genes contain multiple copies of G-C rich repeats and are devoid of TATA boxes. These G-C nonamers represent a motif that may function as a canonical binding site for the transcription factor Sp1 [202]. Mapping of PrP genes to the short arm of Hu chromosome 20 and to the homologous region of Mo chromosome 2 argues for the existence of PrP genes prior to the speciation of mammals [203, 204].

Although PrP mRNA is constitutively expressed in the brains of adult animals [83, 84], it is highly regulated during development. In the septum, levels of PrP mRNA and choline acetyltransferase were found to increase in parallel during development [205]. In other brain regions, PrP gene expression occurred at an earlier age. *In situ* hybridization studies show that the highest levels of PrP mRNA are found in neurons [206].

PrP^C expression in brain was defined by standard immunohistochemistry [207] and by histoblotting in the brains of uninfected controls [208]. Immunostaining of PrP^C in the SHa brain was most intense in the stratum radiatum and stratum oriens of the CA1 region of the hippocampus and was virtually absent from the granule cell layer of the dentate gyrus and the pyramidal cell layer throughout Ammon's horn. PrP^{Sc} staining was minimal in those regions which were intensely stained for PrP^C. A similar relationship between PrP^C and PrP^{Sc} was found in the amygdala. In contrast, PrP^{Sc} accumulated in the medial habenular nucleus, the medial septal nuclei, and the diagonal band of Broca; in contrast, these areas were virtually devoid of PrP^C. In the white matter, bundles of myelinated axons contained PrP^{Sc} but were devoid of PrP^C. These findings suggest that prions are transported along axons and are in agreement with earlier findings in which scrapie infectivity was found to migrate in a pattern consistent with retrograde transport [209–211].

Molecular genetics of prion diseases. Independent of enriching brain fractions for scrapie infectivity that led to the discovery of PrP^{Sc}, the PrP gene was shown to be genetically linked to a locus controlling scrapie incubation times [212]. Subsequently, mutation of the PrP gene was shown to be genetically linked to the development of familial prion disease [4]. At the same time, expression of a SHaPrP transgene in mice was shown to render the animals highly susceptible to SHa prions demonstrating that expression of a foreign PrP gene could abrogate the species barrier [213]. Later, PrP-deficient (Prnp$^{0/0}$) mice were found to be resistant to prion infection and failed to replicate prions, as expected [100, 101]. The results of these studies indicated PrP must play a central role in the transmission and pathogenesis of prion disease but equally important, they argued that the abnormal isoform is an essential component of the prion particle [23].

PrP gene dosage controls length of incubation time. Scrapie incubation times in mice were used to distinguish prion strains and to identify a gene controlling its length [135, 214]. This gene was initially called *Sinc* based on genetic crosses between C57Bl and VM mice that exhibited short and long incubation times, respectively [214]. Because of the restricted distribution of VM mice, we searched for another mouse with long incubation times. I/Ln mice proved to be a suitable substitute for VM mice [215]; eventually, I/Ln and VM mice were

found to be derived from a common ancestor [216]. With a PrP cDNA probe, we demonstrated genetic linkage between the PrP gene and a locus controlling the incubation time in crosses between NZW/LacJ and I/Ln mice [212]. We provisionally designated these genes as components of the *Prn* complex but eventually found that the incubation time gene, *Prn-i*, is either congruent with or closely linked to the PrP gene, *Prnp* [194].

Although the amino acid substitutions in PrP that distinguish NZW *(Prnp^a)* from I/Ln *(Prnp^b)* mice argued for congruency of *Prnp* and *Prn-i*, experiments with *Prnp^a* mice expressing *Prnp^b* transgenes demonstrated a "paradoxical" shortening of incubation times [195]. We had predicted that these Tg mice would exhibit a prolongation of the incubation time after inoculation with RML prions based on *(Prnp^a* x*Prnp^b)* F1 mice which do exhibit long incubation times. We described those findings as "paradoxical shortening" because we and others had believed for many years that long incubation times are dominant traits [212, 214]. From studies of congenic and transgenic mice expressing different numbers of the *a* and *b* alleles of *Prnp*, we learned that these findings were not paradoxical; indeed, they resulted from increased PrP gene dosage [217]. When the RML isolate was inoculated into congenic and transgenic mice, increasing the number of copies of the *a* allele was found to be the major determinant in reducing the incubation time; however, increasing the number of copies of the *b* allele also reduced the incubation time, but not to the same extent as that seen with the *a* allele. From the foregoing investigations, we concluded that both *Sinc* and *Prn-i* are congruent with PrP [217], and recent gene targeting studies have confirmed this view [218].

Overexpression of wtPrP transgenes. Mice were constructed expressing different levels of the wild-type (wt) SHaPrP transgene [213]. Inoculation of these Tg(SHaPrP) mice with SHa prions demonstrated abrogation of the species barrier resulting in abbreviated incubation times [219]. The length of the incubation time after inoculation with SHa prions was inversely proportional to the level of SHaPrP^C in the brains of Tg(SHaPrP) mice [219]. Bioassays of brain extracts from clinically ill Tg(SHaPrP) mice inoculated with Mo prions revealed that only Mo prions but no SHa prions were produced. Conversely, inoculation of Tg(SHaPrP) mice with SHa prions led only to the synthesis of SHa prions. Although the rate of PrP^{Sc} synthesis appears to be a function of the level of PrP^C expression in Tg mice, the level to which PrP^{Sc} finally accumulates seems to be independent of PrP^C concentration [219].

During transgenetic studies, we discovered that uninoculated older mice harboring numerous copies of wtPrP transgenes derived from Syrian hamsters, sheep, and *Prnp^b* mice spontaneously developed truncal ataxia, hindlimb paralysis, and tremors [197]. These Tg mice exhibited a profound necrotizing myopathy involving skeletal muscle, a demyelinating polyneuropathy, and focal vacuolation of the CNS. Development of disease was dependent on transgene dosage. For example, Tg(SHaPrP$^{+/+}$)7 mice homozygous for the SHaPrP transgene array regularly developed disease between 400 and 600 days of age, while hemizygous Tg(SHaPrP$^{+/0}$)7 mice also developed disease, but only after >650 days.

PrP-deficient mice. The development and lifespan of two lines of PrP-deficient (Prnp$^{0/0}$) mice were indistinguishable from controls [220, 221] while two other lines exhibited ataxia and Purkinje cell degeneration at ~70 weeks of age [222] (R. Moore and D. Melton, in preparation). In the former two lines with normal development, altered sleep wake cycles have been reported [223] and altered synaptic behavior in brain slices was reported [224, 225] but could not be confirmed by others [226, 227].

Prnp$^{0/0}$ mice are resistant to prions [100, 101]. Prnp$^{0/0}$ mice were sacrificed 5, 60, 120, and 315 days after inoculation with RML prions and brain extracts bioassayed in CD-1 Swiss mice. Except for residual infectivity from the inoculum detected at 5 days after inoculation, no infectivity was detected in the brains of Prnp$^{0/0}$ mice [101]. One group of investigators found that Prnp$^{0/0}$ mice inoculated with RML prions and sacrificed 20 weeks later had $10^{3.6}$ ID$_{50}$ units/ml of homogenate by bioassay [100]. Others have used this report to argue that prion infectivity replicates in the absence of PrP [67, 132]. Neither we nor the authors of the initial report could confirm the finding of prion replication in Prnp$^{0/0}$ mice [101, 103].

Prion species barrier and protein X. The passage of prions between species is often a stochastic process, almost always characterized by prolonged incubation times during the first passage in the new host [36]. This prolongation is often referred to as the "species barrier" [36, 228]. Prions synthesized *de novo* reflect the sequence of the host PrP gene and not that of the PrP^{Sc} molecules in the inoculum derived from the donor [90]. On subsequent passage in a homologous host, the incubation time shortens to that recorded for all subsequent passages. From studies with Tg mice, three factors have been identified that contribute to the species barrier: (i) the difference in PrP sequences between the prion donor and recipient, (ii) the strain of prion, and (iii) the species specificity of protein X, a factor defined by molecular genetic studies that binds to PrP^C and facilitates PrP^{Sc} formation. This factor is likely to be a protein, hence the provisional designation protein X [134, 178]. The prion donor is the last mammal in which the prion was passaged and its PrP sequence represents the "species" of the prion. The strain of prion, which seems to be enciphered in the conformation of PrP^{Sc}, conspires with the PrP sequence, which is specified by the recipient, to determine the tertiary structure of nascent PrP^{Sc}. These principles are demonstrated by studies on the transmission of Syrian hamster (SHa) prions to mice showing that expression of a SHaPrP transgene in mice abrogated the species barrier (Table 3) [213]. Besides the PrP sequence, the strain of prion modified transmission of SHa prions to mice (Table 3) [135, 229, 230].

Transmission of human prions. Protein X was postulated to explain the results on the transmission of human (Hu) prions to Tg mice (Table 4) [134, 179]. Mice expressing both Mo and HuPrP were resistant to Hu prions while those expressing only HuPrP were susceptible. These results argue that MoPrP^C inhibited transmission of Hu prions i.e. the formation of nascent HuPrP^{Sc}. In contrast to the foregoing studies, mice expressing both MoPrP and chimeric MHu2M PrP were susceptible to Hu prions and mice expressing MHu2MPrP

Table 3. Influence of prion species and strains on transmission across a species barrier*

Inoculum	Host	Sc237		139H	
		Incubation time / [days ± SEM] (n/n$_o$)			
SHa	SHa	77 ± 1	(48/48)	167 ± 1	(94/94)
SHa	non-Tg mice	>700	(0/9)	499 ± 15	(11/11)
SHa	Tg(SHaPrP)81/ FVB mice	75 ± 2	(22/22)	110 ± 2	(19/19)
SHa	Tg(SHaPrP)81/ Prnp$^{0/0}$ mice	54 ± 1	(9/9)	65 ± 1	(15/15)

*data from references [101, 219, 230] (D. Groth and S.B. Prusiner, unpublished data). n/n$_o$ = number of diseased animals / number of injected animals.

alone were only slightly more susceptible. These findings contend that MoPrP^C has only a minimal effect on the formation of chimeric MHu2MPrP^{Sc}.

Table 4. Evidence for protein X from transmission studies of human prions†

Inoculum	Host	MoPrP gene	Incubation Time	
			[days ± SEM] (n/no)	
sCJD	Tg(HuPrP)	Prnp$^{+/+}$	721	(1/10)
sCJD	Tg(HuPrP)Prnp$^{0/0}$	Prnp$^{0/0}$	263 ± 2	(6/6)
sCJD	Tg(MHu2M)	Prnp$^{+/+}$	238 ± 3	(8/8)
sCJD	Tg(MHu2M)Prnp$^{0/0}$	Prnp$^{0/0}$	191 ± 3	(10/10)

†data with inoculum RG [134].

Genetic evidence for protein X. When the data on Hu prion transmission to Tg mice were considered together, they suggested that MoPrP^C prevented the conversion of HuPrP^C into PrP^{Sc} but had little effect on the conversion of MHu2M into PrP^{Sc} by binding to another Mo protein. We interpreted these results in terms of MoPrP^C binding to this Mo protein with a higher affinity than does HuPrP^C. We postulated that MoPrP^C had little effect on the formation of PrP^{Sc} from MHu2M (Table 4) because MoPrP and MHu2M share the same amino acid sequence at the COOH-terminus. We hypothesized that MoPrP^C only weakly inhibited transmission of SHa prions to Tg(SHaPrP) mice (Table 3) because SHaPrP is more closely related to MoPrP than is HuPrP.

Using scrapie-infected mouse (Mo) neuroblastoma cells transfected with chimeric Hu/Mo PrP genes, we extended our studies of protein X. Substitution of a Hu residue at position 214 or 218 prevented PrP^{Sc} formation (Figure 5B) [178]. The side chains of these residues protrude from the same surface of the COOH-terminal α-helix forming a discontinuous epitope with residues 167 and 171 in an adjacent loop. Substitution of a basic residue at positions 167, 171, or 218 prevented PrP^{Sc} formation; these mutant PrPs appear to act as "dominant negatives" by binding protein X and rendering it unavailable for prion propagation. Our findings within the context of protein X explain the protective effects of basic polymorphic residues in PrP of humans and sheep [198, 231, 232].

Is protein X a molecular chaperone? Since PrP undergoes a profound structural transition during prion propagation, it seems likely that other proteins

such as chaperones participate in this process. Whether protein X functions as a molecular chaperone is unknown. Interestingly, scrapie-infected cells in culture display marked differences in the induction of heat-shock proteins [233, 234] and Hsp70 mRNA has been reported to increase in scrapie of mice [235]. While attempts to isolate specific proteins that bind to PrP have been disappointing [236], PrP has been shown to interact with Bcl-2 and Hsp60 by two-hybrid analysis in yeast [237, 238]. Although these studies are suggestive, no molecular chaperone involved in prion formation in mammalian cells has been identified.

Miniprions. Using the four-helix bundle model of PrP^C (Figure 4A) [147], each region of proposed secondary structure was systematically deleted and the mutant constructs expressed in scrapie-infected neuroblastoma (ScN2a) cells and Tg mice [164, 168]. Deletion of any of the four putative helical regions prevented PrP^{Sc} formation while deletion of the NH$_2$-terminal region containing residues 23–89 did not affect the yield of PrP^{Sc}. In addition to the 67 residues at the NH$_2$-terminus, 36 residues from position 141 to 176 could be deleted without altering PrP^{Sc} formation (Figures 6 and 7). The resulting PrP molecule of 106 amino acids was designated PrP106. In this mutant PrP, helix A as well as the S2 β-strand were removed. When PrP106 was expressed in ScN2a cells, PrP^{Sc}106 was soluble in 1% Sarkosyl. Whether the structure of PrP^{Sc}106 can be more readily determined than that of full-length PrP^{Sc} remains uncertain.

Transgene-specified susceptibility. Tg(PrP106)Prnp$^{0/0}$ mice that expressed PrP106 developed neurological dysfunction ~300 days after inoculation with RML prions previously passaged in CD-1 Swiss mice (S. Supattapone, T. Muramoto, D. Peretz, S. J. DeArmond, A. Wallace, F. E. Cohen, S. B. Prusiner and M. R. Scott, in preparation). The resulting prions containing PrP^{Sc}106

PrP(90-231)
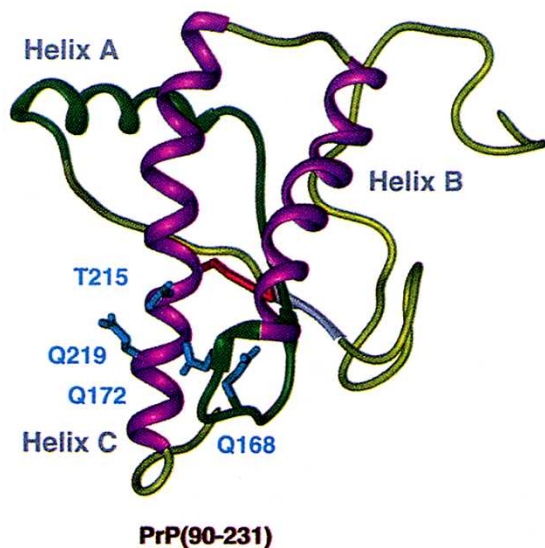
Figure 6. Miniprions produced by deleting PrP residues 23–89 and 141–176. The deletion of residues 141–176 (green) containing helix A and the S2 β-strand is shown.
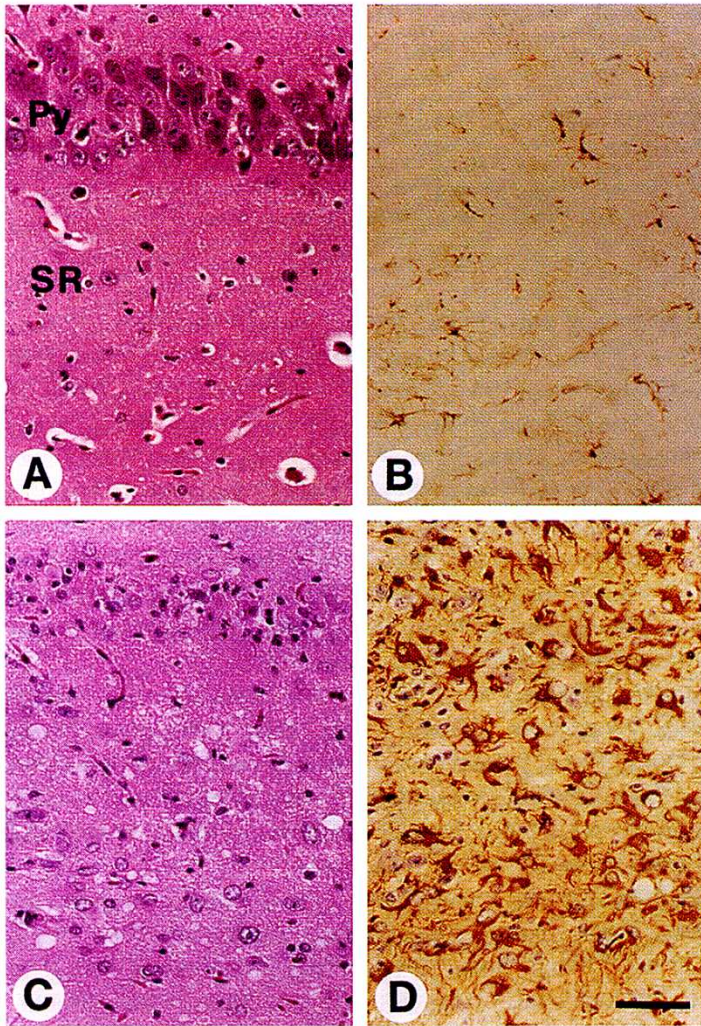
Figure 7. Tg(PrP106) Prnp$^{0/0}$ mice were inoculated with RML106 prions containing PrP^{Sc}106. Sections of the hippocampus stained with hematoxylin (A and C) and eosin (B and D). (A and B) Control Tg(PrP106) mouse uninoculated and without neurologic deficits. (C and D) Tg(PrP106) Prnp$^{0/0}$ mouse inoculated with RML106 prions and sacrificed after signs of neurologic dysfunction observed. Photomicrographs prepared by Stephen J. DeArmond.

produced CNS disease in ~66 days upon subsequent passage in Tg(PrP106)Prnp$^{0/0}$ mice. (Table 5). Besides widespread spongiform degeneration and PrP deposits, the pyramidal cells of the hippocampus comprising the CA-1, CA-2, and CA-3 fields disappeared in Tg(PrP106)Prnp$^{0/0}$ mice inoculated with prions containing PrP^{Sc}106 (Figure 7A–D). In no previous study of Tg mice have we seen similar neuropathological lesions. The Tg(MoPrP-A) mice overexpressing MoPrP are resistant to RML106 miniprions, but are highly susceptible to RML prions. These mice require more than 250 days to produce illness after inoculation with miniprions but develop disease in ~50 days when inoculated with RML prions containing full-length MoPrP^{Sc}.

Table 5. Susceptibility and resistance of transgenic mice to artificial miniprions*

Host	RML106 miniprions		RML prions	
	Incubation time [days ± SEM] (n/n_o)			
Tg(PrP106)Prnp$^{0/0}$ mice	66 ± 3	(10/10)	300 ± 2	(9/10)
Tg(MoPrP-A) mice	>250	(0/11)	50 ± 2	(16/16)

*data from references [217] (S. Supattapone, T. Muramoto, D. Peretz, S. J. DeArmond, A. Wallace, F. E. Cohen, S. B. Prusiner and M. R. Scott, in preparation).

Smaller prions and mythical viruses. The unique incubation times and neuropathogy in Tg mice caused by miniprions are difficult to reconcile with the notion that scrapie is caused by an as yet unidentified virus. When the mutant or wt PrP^C of the host matched PrP^{Sc} in the inoculum, the mice were highly susceptible (Table 5). However, when there was a mismatch between PrP^C and PrP^{Sc}, the mice were resistant to the prions. This principle of homologous PrP interactions which underlies the species barrier (Table 3) is recapitulated in studies of PrP106 where the amino acid sequence has been drastically changed by deleting nearly 50% of the residues. Indeed, the unique properties of the miniprions provide another persuasive argument supporting the contention that prions are infectious proteins.

Human prion diseases. Most humans afflicted with prion disease present with rapidly progressive dementia, but some manifest cerebellar ataxia. Although the brains of patients appear grossly normal upon post-mortem examination, they usually show spongiform degeneration and astrocytic gliosis under the microscope. The human prion diseases can present as sporadic, genetic, or infectious disorders [5] (Table 1A).

Sporadic CJD. Sporadic forms of prion disease comprise most cases of CJD and possibly a few cases of Gerstmann-Sträussler-Scheinker disease (GSS) (Table 1A) [4, 239, 240]. In these patients, mutations of the PrP gene are not found. How prions causing disease arise in patients with sporadic forms is unknown; hypotheses include horizontal transmission of prions from humans or animals [241], somatic mutation of the PrP gene, and spontaneous conversion of PrP^C into PrP^{Sc} [5, 15]. Since numerous attempts to establish an infectious link between sporadic CJD and a preexisting prion disease in animals or humans have been unrewarding, it seems unlikely that transmission features in the pathogenesis of sporadic prion disease [9–12, 242].

Inherited prion diseases. To date, 20 different mutations in the human PrP gene resulting in nonconservative substitutions have been found that segregate with the inherited prion diseases (Figure 4B). Familial CJD cases suggested that genetic factors might influence pathogenesis [1, 2, 243], but this was difficult to reconcile with the transmissibility of fCJD and GSS [3]. The discovery of genetic linkage between the PrP gene and scrapie incubation times in mice [212] raised the possibility that mutation might feature in the hereditary human prion diseases. The P102L mutation was the first PrP mutation to be genetically linked to CNS dysfunction in GSS (Figure 4B)[4] and has since been found in many GSS families throughout the world [244-246]. Indeed, a mutation

in the protein coding region of the PrP gene has been found in all reported kindred with familial human prion disease; besides the P102L mutation, genetic linkage has been established for four other mutations [16, 29–31].

Tg mouse studies confirmed that mutations of the PrP gene can cause neurodegeneration. The P102L mutation of GSS was introduced into the MoPrP transgene and five lines of Tg(MoPrP-P101L) mice expressing high levels of mutant PrP developed spontaneous CNS degeneration consisting of widespread vacuolation of the neuropil, astrocytic gliosis, and numerous PrP amyloid plaques similar to those seen in the brains of humans who die from GSS(P102L) [247–249]. Brain extracts prepared from spontaneously ill Tg(MoPrP-P101L) mice transmitted CNS degeneration to Tg196 mice but contained no protease-resistant PrP [248, 249]. The Tg196 mice do not develop spontaneous disease but express low levels of the mutant transgene MoPrP-P101L and are deficient for mouse PrP (Prnp$^{0/0}$) [220]. These studies, combined with the transmission of prions from patients who died of GSS to apes and monkeys [3] or to Tg(MHu2M-P101L) mice [134], demonstrate that prions are generated de novo by mutations in PrP. Additionally, brain extracts from patients with some other inherited prion diseases, fCJD(E200K) or FFI, transmit disease to Tg(MHu2M) mice [27]. An artificial set of mutations in a PrP transgene consisting of A113V, A115V, and A118V produced neurodegeneration in neonatal mice; these Val substitutions were selected for their propensity to form β sheets [153, 250]. In preliminary studies, brain extracts from two of these mice transmitted disease to hamsters and to Tg mice expressing a chimeric SHa/Mo PrP.

Genetic disease that is transmissible. Had the PrP gene been identified in families with prion disease by positional cloning or through the purification and sequencing of PrP in amyloid plaques before brain extracts were shown to be transmissible, the prion concept might have been more readily accepted. Within that scenario, it seems likely that we would have explored the possibility that the mutant protein, upon inoculation into a susceptible host, stimulated production of more of a similar protein. Postulating an infectious pathogen with a foreign genome would have been the least likely candidate to explain how a genetic disease could be experimentally transmissible.

Infectious prion diseases. The infectious prion diseases include kuru of the Fore people in New Guinea where prions were transmitted by ritualistic cannibalism [241, 251, 252]. With the cessation of cannibalism at the urging of missionaries, kuru began to decline long before it was known to be transmissible (Figure 8). Sources of prions causing infectious CJD on several different continents include improperly sterilized depth electrodes, transplanted corneas, human growth hormone (HGH) and gonadotrophin derived from cadaveric pituitaries, and dura mater grafts [253]. Over 90 young adults have developed CJD after treatment with cadaveric HGH; the incubation periods range from 3 to more than 20 years [254, 255]. Dura mater grafts implanted during neurosurgical procedures seem to have caused more than 60 cases of CJD; these incubation periods range from one to more than 14 years [256–258].

Prion diversity. The existence of prion strains raises the question of how
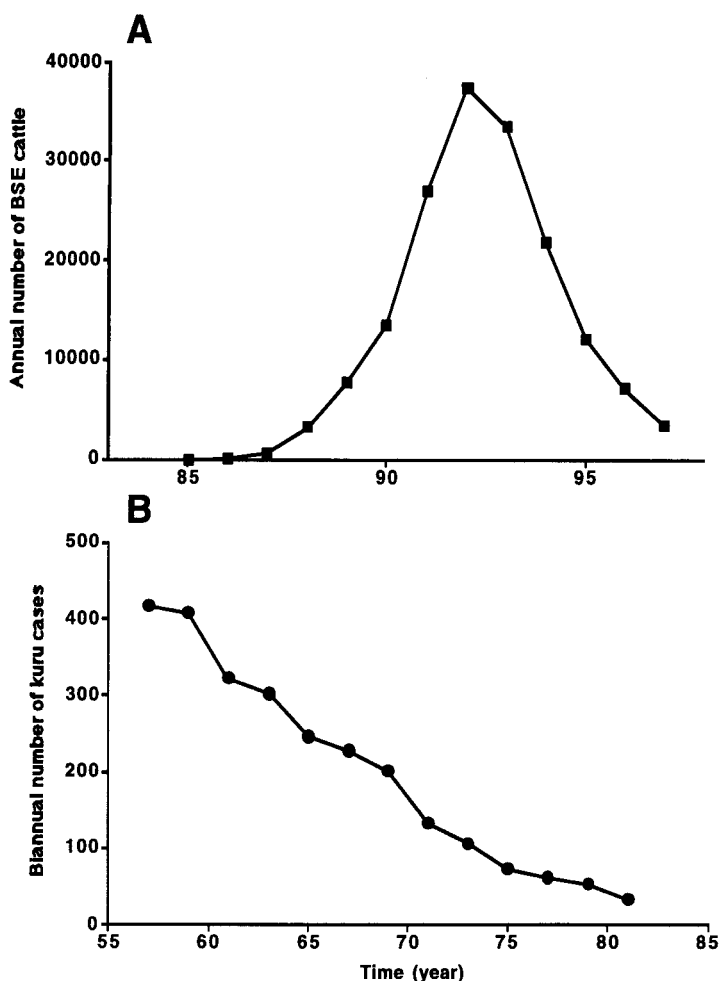
Figure 8. Disappearance of the kuru and BSE epidemics. (A) Number of annual cases of BSE in cattle in Great Britain. (B) Number of biannual cases of kuru in Papua New Guinea. Data compiled for BSE by John Wilesmith and for kuru by Michael Alpers.

heritable biological information can be enciphered in a molecule other than nucleic acid [131, 214, 259-263]. Strains or varieties of prions have been defined by incubation times and the distribution of neuronal vacuolation [214, 264]. Subsequently, the patterns of PrP^{Sc} deposition were found to correlate with vacuolation profiles and these patterns were also used to characterize strains of prions [230, 265, 266]

The typing of prion strains in C57Bl, VM, and F1(C57Bl x VM) inbred mice began with isolates from sheep with scrapie. The prototypic strains called Me7 and 22A gave incubation times of ~150 and ~400 days in C57Bl mice, respectively [214, 267, 268]. The PrP genes of C57Bl and I/Ln (and later VM) mice encode proteins differing at two residues and control scrapie incubation times [194, 212, 216–218, 269].

Until recently, support for the hypothesis that the tertiary structure of PrP^{Sc}

enciphers strain-specific information [23] was minimal except for the DY strain isolated from mink with transmissible encephalopathy by passage in Syrian hamsters [26, 270, 271]. PrP^{Sc} in DY prions showed diminished resistance to proteinase K digestion as well as a peculiar site of cleavage. The DY strain presented a puzzling anomaly since other prion strains exhibiting similar incubation times did not show this altered susceptibility to proteinase K digestion of PrP^{Sc} [135]. Also notable was the generation of new strains during passage of prions through animals with different PrP genes [135, 229].

PrP^{Sc} conformation enciphers diversity. Persuasive evidence that strain-specific information is enciphered in the tertiary structure of PrP^{Sc} comes from transmission of two different inherited human prion diseases to mice expressing a chimeric MHu2M PrP transgene [27]. In fatal familial insomnia (FFI), the protease-resistant fragment of PrP^{Sc} after deglycosylation has an M_r of 19 kD; whereas in fCJD(E200K) and most sporadic prion diseases, it is 21 kD (Table 6) [272, 273]. This difference in molecular size was shown to be due to different sites of proteolytic cleavage at the NH$_2$-termini of the two human PrP^{Sc} molecules reflecting different tertiary structures [272]. These distinct conformations were understandable since the amino acid sequences of the PrPs differ.

Table 6. Distinct prion strains generated in humans with inherited prion diseases and transmitted to transgenic mice*

Inoculum	Host Species	Host PrP Genotype	Incubation time [days ± SEM] (n/n_o)		PrP^{Sc}(kD)
None	Human	FFI(D178N,M129)	19		
FFI	Mouse	Tg(MHu2M)	206 ± 7	(7/7)	19
FFI → Tg(MHu2M)	Mouse	Tg(MHu2M)	136 ± 1	(6/6)	19
None	Human	fCJD(E200K)	21		
fCJD	Mouse	Tg(MHu2M)	170 ± 2	(10/10)	21
fCJD → Tg(MHu2M)	Mouse	Tg(MHu2M)	167 ± 3	(15/15)	21

*data from reference [27] (G. Telling *et al.*, in preparation).

Extracts from the brains of FFI patients transmitted disease to mice expressing a chimeric MHu2M PrP gene about 200 days after inoculation and induced formation of the 19 kD PrP^{Sc}; whereas fCJD(E200K) and sCJD produced the 21 kD PrP^{Sc} in mice expressing the same transgene [27]. On second passage, Tg(MHu2M) mice inoculated with FFI prions showed an incubation time of ~130 days and a 19 kD PrP^{Sc} while those inoculated with fCJD(E200K) prions exhibited an incubation time of ~170 days and a 21 kD PrP^{Sc} [28]. The experimental data demonstrate that MHu2MPrP^{Sc} can exist in two different conformations based on the sizes of the protease-resistant fragments; yet, the amino acid sequence of MHu2MPrP^{Sc} is invariant.

The results of our studies argue that PrP^{Sc} acts as a template for the conversion of PrP^{C} into nascent PrP^{Sc}. Imparting the size of the protease-resistant fragment of PrP^{Sc} through conformational templating provides a mechanism for both the generation and propagation of prion strains.

Interestingly, the protease-resistant fragment of PrP^{Sc} after deglycosylation with an M_r of 19 kD has been found in a patient who developed a sporadic

case of prion disease similar to FFI but with no family history. Since both PrP alleles encoded the wt sequence and a Met at position 129, we labeled this case, fatal sporadic insomnia (FSI). At autopsy, the spongiform degeneration, reactive astrogliosis, and PrP[Sc] deposition were confined to the thalamus [274]. These findings argue that the clinicopathologic phenotype is determined by the conformation of PrP[Sc] in accord with the results of the transmission of human prions from patients with FFI to Tg mice [27].

Selective neuronal targeting. Besides incubation times, profiles of spongiform change (Figure 1) have been used to characterize prion strains [275], but recent studies argue that such profiles are not an intrinsic feature of strains [276, 277]. The mechanism by which prion strains modify the pattern of spongiform degeneration was perplexing since earlier investigations had shown that PrP[Sc] deposition precedes neuronal vacuolation and reactive gliosis [211, 230]. When FFI prions were inoculated into Tg(MHu2M) mice, PrP[Sc] was confined largely to the thalamus (Figure 9A) as is the case for FFI in humans [27, 278]. In contrast, fCJD(E200K) prions inoculated into Tg(MHu2M) mice produced widespread deposition of PrP[Sc] throughout the cortical mantle and many of the deep structures of the CNS (Figure 9B) as is seen in fCJD(E200K) of humans. To examine whether the diverse patterns of PrP[Sc] deposition are influenced by Asn-linked glycosylation of PrP[C], we constructed Tg mice expressing PrPs mutated at one or both of the Asn-linked glycosylation consensus sites [277]. These mutations resulted in aberrant neuroanatomic topologies of PrP[C] within the CNS, whereas pathologic point mutations adjacent to the consensus sites did not alter the distribution of PrP[C]. Tg mice with mutation of the second PrP glycosylation site exhibited prion incubation times of >500 days and unusual patterns of PrP[Sc] deposition. These findings raise two possible scenarios. First, glycosylation can modify the conformation of PrP[C] and affect its affinity for a particular conformer of PrP[Sc], which results in specific patterns of PrP[Sc] deposition; such interactions between PrP[Sc] and PrP[C] are likely to determine the rate of nascent PrP[Sc] formation. Second, glycosylation modifies the stability of PrP[Sc] and hence, the rate of PrP[Sc] clearance. This latter explanation is consistent with the proposal that the binding of PrP[C] to protein X is the rate-limiting step in PrP[Sc] formation under most circumstances [255].

Bovine spongiform encephalopathy. Prion strains and the species barrier are of paramount importance in understanding the BSE epidemic in Great Britain, in which it is estimated that almost one million cattle were infected with prions [279, 280]. The mean incubation time for BSE is about 5 years. Most cattle therefore did not manifest disease since they were slaughtered between 2 and 3 years of age [281]. Nevertheless, more than 160,000 cattle, primarily dairy cows, have died of BSE over the past decade (Figure 5A) [279]. BSE is a massive common source epidemic caused by meat and bone meal (MBM) fed primarily to dairy cows [280, 282]. The MBM was prepared from the offal of sheep, cattle, pigs, and chickens as a high protein nutritional supplement. In the late 1970s, the hydrocarbon-solvent extraction method used in the rendering of offal began to be abandoned resulting in MBM with a much higher
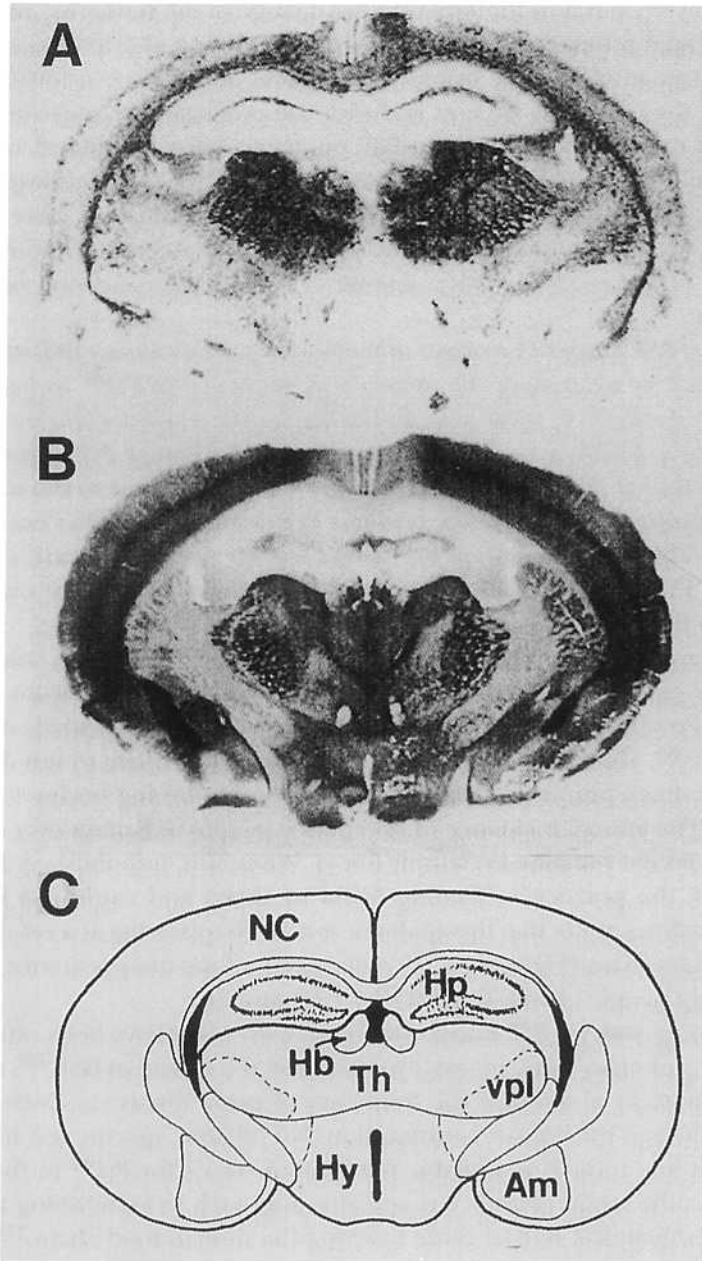
Figure 9. Regional distribution of PrP^{Sc} deposition in Tg(MHu2M)Prnp$^{0/0}$ mice inoculated with prions from humans who died of inherited prion diseases (Table 5). Histoblot of PrP^{Sc} deposition in a coronal section a Tg(MHu2M)Prnp$^{0/0}$ mouse through the hippocampus and thalamus [27]. (A) The Tg mouse was inoculated with brain extract prepared from a patient who died of FFI. (B) The Tg mouse was inoculated with extract from a patient with fCJD(E200K). Cryostat sections were mounted on nitrocellulose and treated with proteinase K to eliminate PrP^C [208]. To enhance the antigenicity of PrP^{Sc}, the histoblots were exposed to 3 guanidinium isothiocyanate before immunostaining using anti-PrP 3F4 mAb [174]. (C) Labelled diagram of a coronal sections of the hippocampus/thalamus region. NC, neocortex; Hp, hippocampus; Hb, habenula; Th, thalamus; vpl, ventral posterior lateral thalamic nucleus; Hy, hypothalamus; Am, amygdala. Photomicrographs prepared by Stephen J. DeArmond.

fat content [282]. It is now thought that this change in the rendering process allowed scrapie prions from sheep to survive rendering and to be passed into cattle. Alternatively, bovine prions were present at low levels prior to modification of the rendering process and with the processing change survived in sufficient numbers to initiate the BSE epidemic when inoculated back into cattle orally through MBM. Against the latter hypothesis is the widespread geographical distribution throughout England of the initial 17 cases of BSE, which occurred almost simultaneously [280, 283, 284]. Furthermore, there is no evidence of a pre-existing prion disease of cattle, either in Great Britain or elsewhere.

Origin of BSE prions? The origin of the bovine prions causing BSE cannot be determined by examining the amino acid sequence of PrP^{Sc} in cattle with BSE since the PrP^{Sc} in these animals has the bovine sequence whether the initial prions in MBM came from cattle or sheep. The bovine PrP sequence differs from that of sheep at 7 or 8 positions [285–287]. In contrast to the many PrP polymorphisms found in sheep, only one PrP polymorphism has been found in cattle. Though most bovine PrP alleles encode five octarepeats, some encode six. PrP alleles encoding six octarepeats do not seem to be overrepresented in BSE (Figure 4B) [288].

Brain extracts from BSE cattle cause disease in cattle, sheep, mice, pigs, and mink after intracerebral inoculation [289–293], but prions in brain extracts from sheep with scrapie fed to cattle produced illness substantially different from BSE [294]. However, no exhaustive effort has been made to test different strains of sheep prions or to examine the disease following bovine to bovine passage. The annual incidence of sheep with scrapie in Britain over the past two decades has remained relatively low (J. Wilesmith, unpublished data). In July 1988, the practice of feeding MBM to sheep and cattle was banned. Recent statistics argue that the epidemic is now disappearing as a result of this ruminant feed ban (Figure 8A) [279], reminiscent of the disappearance of kuru in the Fore people of New Guinea [241, 252] (Figure 8B).

Monitoring cattle for BSE prions. Although many plans have been offered for the culling of older cattle in order to minimize the spread of BSE [279], it seems more important to monitor the frequency of prion disease in cattle as they are slaughtered for human consumption. No reliable, specific test for prion disease in live animals is available but immunoassays for PrP^{Sc} in the brainstems of cattle might provide a reasonable approach to establishing the incidence of subclinical BSE in cattle entering the human food chain [176, 208, 287, 295–297]. Determining how early in the incubation period PrP^{Sc} can be detected by immunological methods is now possible since a reliable bioassay has been created by expressing the BoPrP gene in Tg mice [298]. Prior to development of Tg(BoPrP)Prnp$^{0/0}$ mice, non-Tg mice inoculated intracerebrally with BSE brain extracts required more than 300 days to develop disease [67, 293, 299, 300]. Depending on the titer of the inoculum, the structures of PrP^C and PrP^{Sc}, and the structure of protein X, the number of inoculated animals developing disease can vary over a wide range (Table 3). Some investigators have stated that transmission of BSE to mice is quite variable with incubation periods ex-

ceeding one year [67] while others report low prion titers in BSE brain homogenates [299, 300] compared to rodent brain scrapie [54, 56, 301, 302].

Have bovine prions been transmitted to humans? In 1994, the first cases of CJD in teenagers and young adults that were eventually labelled new variant (v) CJD occurred in Britain [303]. Besides the young age of these patients [304, 305], the brains of these patients showed numerous PrP amyloid plaques surrounded by a halo of intense spongiform degeneration (Figure 10A and B) [306]. Later, one French case meeting these criteria followed [307]. These unusual neuropathologic changes have not been seen in CJD cases in the United States, Australia, or Japan [306, 308]. Both macaque monkeys and marmosets developed neurologic disease several years after inoculation with bovine
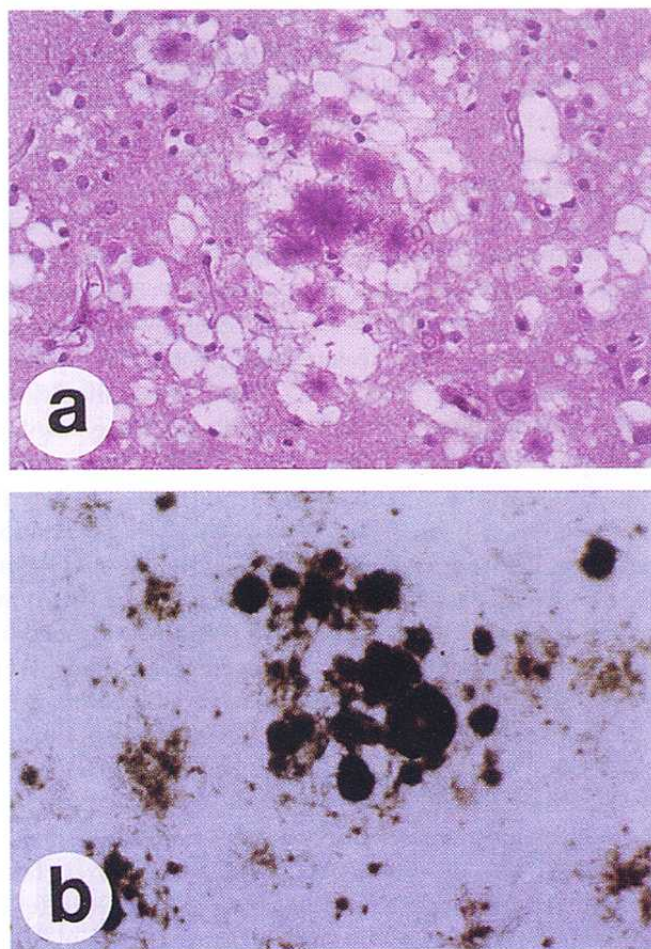
Figure 10. Histopathology of variant Creutzfeldt-Jakob disease in Great Britain. (A) Section from frontal cortex stained by the periodic acid Schiff method showing a field with aggregates of plaques surrounded by spongiform degeneration. Magnification X 100. (B) Multiple plaques and amorphous deposits are PrP immunopositive. Magnification X 500. Specimens provided by James Ironside, Jeanne Bell, and Robert Will; photomicrographs prepared by Stephen J. DeArmond.

prions [309], but only the macaques exhibited numerous PrP plaques similar to those found in vCJD [310] (R. Ridley and H. Baker, unpublished data).

The restricted geographical occurrence and chronology of vCJD have raised the possibility that BSE prions have been transmitted to humans. That only ~25 vCJD cases have been recorded and the incidence has remained relatively constant makes establishing the origin of vCJD difficult. No set of dietary habits distinguishes vCJD patients from apparently healthy people. Moreover, there is no explanation for the predilection of vCJD for teenagers and young adults. Why have older individuals not developed vCJD-based neuropathologic criteria? It is noteworthy that epidemiological studies over the past three decades have failed to find evidence for transmission of sheep prions to humans [9–12]. Attempts to predict the future number of cases of vCJD, assuming exposure to bovine prions prior to the offal ban, have been uninformative because so few cases of vCJD have occurred [311–313]. Are we at the beginning of a human prion disease epidemic in Britain like those seen for BSE and kuru (Figure 8) or will the number of vCJD cases remain small as seen with iCJD caused by cadaveric HGH [254, 255]?

Strain of BSE prions. Was a particular conformation of bovine PrP^{Sc} selected for heat-resistance during the rendering process and then reselected multiple times as cattle infected by ingesting prion-contaminated MBM were slaughtered and their offal rendered into more MBM? Recent studies of PrP^{Sc} from brains of patients who died of vCJD show a pattern of PrP glycoforms different from those found for sCJD or iCJD [314, 315]. But the utility of measuring PrP glycoforms is questionable in trying to relate BSE to vCJD [316, 317] because PrP^{Sc} is formed after the protein is glycosylated [138, 140] and enzymatic deglycosylation of PrP^{Sc} requires denaturation [318, 319]. Alternatively, it may be possible to establish a relationship between the conformations of PrP^{Sc} from cattle with BSE and those from humans with vCJD by using Tg mice as was done for strains generated in the brains of patients with FFI or fCJD [27, 298]. A relationship between vCJD and BSE has been suggested by finding similar incubation times in non-Tg RIII mice of ~310 days after inoculation with Hu or Bo prions [293].

Yeast and other prions. Although prions were originally defined in the context of an infectious pathogen [58], it is now becoming widely accepted that prions are elements that impart and propagate variability through multiple conformers of a normal cellular protein. Such a mechanism must surely not be restricted to a single class of transmissible pathogens. Indeed, it is likely that the original definition will need to be extended to encompass other situations where a similar mechanism of information transfer occurs.

Two notable prion-like determinants, [URE3] and [PSI], have already been described in yeast and one in another fungus denoted [Het-s*] [320–325]. Studies of candidate prion proteins in yeast may prove particularly helpful in the dissection of some of the events that feature in PrP^{Sc} formation. Interestingly, different strains of yeast prions have been identified [326]. Conversion to the prion-like [PSI] state in yeast requires the molecular chaperone Hsp104; however, no homolog of Hsp104 has been found in mammals [321, 327]. The NH$_2$-terminal prion domains of Ure2p and Sup35 that are

responsible for the [URE3] and [PSI] phenotypes in yeast have been identified. In contrast to PrP, which is a GPI-anchored membrane protein, both Ure2p and Sup35 are cytosolic proteins [328]. When the prion domains of these yeast proteins were expressed in *E. coli*, the proteins were found to polymerize into fibrils with properties similar to those of PrP and other amyloids [322–324].

Whether prions explain some other examples of acquired inheritance in lower organisms is unclear [329, 330]. For example, studies on the inheritance of positional order and cellular handedness on the surface of small organisms have demonstrated the epigenetic nature of these phenomena but the mechanism remains unclear [331, 332].

Prevention of and therapeutics for prion diseases. As our understanding of prion propagation increases, it should be possible to design effective therapeutics. Because people at risk for inherited prion diseases can now be identified decades before neurologic dysfunction is evident, the development of an effective therapy for these fully penetrant disorders is imperative [333, 334]. Although we have no way of predicting the number of individuals who may develop neurologic dysfunction from bovine prions in the future [312], it would be prudent to seek an effective therapy now [28, 335]. Interfering with the conversion of PrP^C into PrP^{Sc} seems to be the most attractive therapeutic target [60]. Either stabilizing the structure of PrP^C by binding a drug or modifying the action of protein X, which might function as a molecular chaperone (Figure 5B), are reasonable strategies. Whether it is more efficacious to design a drug that binds to PrP^C at the protein X binding site or one that mimics the structure of PrP^C with basic polymorphic residues that seem to prevent scrapie and CJD remains to be determined [178, 232]. Since PrP^{Sc} formation seems limited to caveolae-like domains [142–145, 336], drugs designed to inhibit this process need not penetrate the cytosol of cells but they do need to be able to enter the CNS. Alternatively, drugs that destabilize the structure of PrP^{Sc} might also prove useful.

The production of domestic animals that do not replicate prions may also be important with respect to preventing prion disease. Sheep encoding the R/R polymorphism at position 171 seem to be resistant to scrapie [198, 231, 337–343]; presumably, this was the genetic basis of Parry's scrapie eradication program in Great Britain 30 years ago [44, 344]. A more effective approach using dominant negatives for producing prion-resistant domestic animals, including sheep and cattle, is probably the expression of PrP transgenes encoding R171 as well as additional basic residues at the putative protein X binding site (Figure 5B) [178]. Such an approach can be readily evaluated in Tg mice and once shown to be effective, it could be instituted by artificial insemination of sperm from males homozygous for the transgene. More difficult is the production of PrP-deficient cattle and sheep. Although such animals would not be susceptible to prion disease [100, 101], they might suffer some deleterious effects from ablation of the PrP gene [222–224, 227].

Whether gene therapy for the human prion diseases using the dominant negative approach described above for prion-resistant animals will prove fea-

sible depends on the availability of efficient vectors for delivery of the trans-
gene to the CNS.

Concluding remarks–looking to the future. Although the study of prions
has taken several unexpected directions over the past three decades, a rather
novel and fascinating story of prion biology is emerging. Investigations of
prions have elucidated a previously unknown mechanism of disease in hu-
mans and animals. While learning the details of the structures of PrPs and de-
ciphering the mechanism of PrP^C transformation into PrP^{Sc} will be impor-
tant, the fundamental principles of prion biology have become reasonably
clear. Though some investigators prefer to view the composition of the infec-
tious prion particle as unresolved [335, 345], such a perspective denies an enlarg-
ing body of data, none of which refutes the prion concept. Moreover, the dis-
covery of prion-like phenomena mediated by proteins unrelated to PrP in
yeast and fungi serves not only to strengthen the prion concept but also to wi-
den it [328].

Hallmark of prion diseases. The hallmark of all prion diseases–whether
sporadic, dominantly inherited, or acquired by infection–is that they involve
the aberrant metabolism and resulting accumulation of the prion protein
(Table 1)[23]. The conversion of PrP^C into PrP^{Sc} involves a conformation
change whereby the α-helical content diminishes and the amount of β-sheet
increases [25]. These findings provide a reasonable mechanism to explain the
conundrum presented by the three different manifestations of prion disease.

Understanding how PrP^C unfolds and refolds into PrP^{Sc} will be of para-
mount importance in transferring advances in the prion diseases to studies of
other degenerative illnesses. The mechanism by which PrP^{Sc} is formed must
involve a templating process whereby existing PrP^{Sc} directs the refolding of
PrP^C into a nascent PrP^{Sc} with the same conformation. Not only will a knowl-
edge of PrP^{Sc} formation help in the rational design of drugs that interrupt
the pathogenesis of prion diseases, but it may also open new approaches to
deciphering the causes of and to developing effective therapies for the more
common neurodegenerative diseases including Alzheimer's disease, Parkin-
son's disease, and amyotrophic lateral sclerosis (ALS). Indeed, the expanding
list of prion diseases and their novel modes of transmission and pathogenesis
(Table 1), as well as the unprecedented mechanisms of prion propagation
and information transfer (Tables 5 and 6), indicate that much more attention
to these fatal disorders of protein conformation is urgently needed.

But prions may have even wider implications than those noted for the com-
mon neurodegenerative diseases. If we think of prion diseases as disorders of
protein conformation and do not require the diseases to be transmissible,
then what we have learned from the study of prions may reach far beyond
these common illnesses.

Multiple conformers. The discovery that proteins may have multiple biologi-
cally active conformations may prove no less important than the implications
of prions for diseases. How many different tertiary structures can PrP^{Sc}
adopt? This query not only addresses the issue of the limits of prion diversity
but also applies to proteins as they normally function within the cell or act to

affect homeostasis in multicellular organisms. The expanding list of chaperones that assist the folding and unfolding of proteins promises much new knowledge about this process. For example, it is now clear that proproteases can carry their own chaperone activity where the *pro* portion of the protein functions as a chaperone in *cis* to guide the folding of the proteolytically active portion before it is cleaved [346]. Such a mechanism might well feature in the maturation of polypeptide hormones. Interestingly, mutation of the chaperone portion of prosubtilisin resulted in the folding of a subtilisin protease with different properties than the one folded by the wild-type chaperone. Such chaperones have also been shown to work in *trans* [346]. Besides transient metabolic regulation within the cell and hormonal regulation of multicellular organisms, it is not unreasonable to suggest that polymerization of proteins into multimeric structures such as intermediate filaments might be controlled at least in part by alternative conformations of proteins. Such regulation of multimeric protein assemblies might occur either in the proteins that form the polymers or the proteins that function to facilitate the polymerization process. Additionally, apoptosis during development and throughout adult life might also be regulated, at least in part, by alternative tertiary structures of proteins.

Shifting the debate. The debate about prions and the diseases that they cause has now shifted to such issues as how many biological processes are controlled by changes in protein conformation. Although the extreme radiation-resistance of the scrapie infectivity suggested that the pathogen causing this disease and related illnesses would be different from viruses, viroids, and bacteria [32, 33], few thought that alternative protein conformations might even remotely feature in the pathogenesis of the prion diseases [37]. Indeed, an unprecedented mechanism of disease has been revealed where an aberrant conformational change in a protein is propagated. The discovery of prions and their eventual acceptance by the community of scholars represents a triumph of the scientific process over prejudice. The future of this new and emerging area of biology should prove even more interesting and productive as a multitude of unpredicted discoveries emerge.

REFERENCES

1. Meggendorfer F.: Klinische und genealogische Beobachtungen bei einem Fall von spastischer Pseudosklerose Jakobs. *Z. Gesamte Neurol. Psychiatr.* 128:337–341, 1930.
2. Stender A.: Weitere Beiträge zum Kapitel "Spastische Pseudosklerose Jakobs". *Z. Gesamte Neurol. Psychiatr.* 128:528–543, 1930.
3. Masters C.L., Gajdusek D.C., and Gibbs C.J., Jr.: Creutzfeldt-Jakob disease virus isolations from the Gerstmann-Sträussler syndrome. *Brain* 104:559–588, 1981.
4. Hsiao K., Baker H.F., Crow T.J., Poulter M., Owen F., Terwilliger J.D., Westaway D., Ott J., and Prusiner S.B.: Linkage of a prion protein missense variant to Gerstmann-Sträussler syndrome. *Nature* 338:342–345, 1989.
5. Prusiner S.B.: Scrapie prions. *Annu. Rev. Microbiol.* 43:345–374, 1989.
6. Kirschbaum W.R.: *Jakob-Creutzfeldt Disease,* Elsevier, Amsterdam, 1968.
7. Gibbs C.J., Jr., Gajdusek D.C., Asher D.M., Alpers M.P., Beck E., Daniel P.M., and Matthews W.B.: Creutzfeldt-Jakob disease (spongiform encephalopathy): transmission to the chimpanzee. *Science* 161:388–389, 1968.
8. Prusiner S.B.: The prion diseases. *Sci. Am.* 272:48–57, 1995.
9. Malmgren R., Kurland L., Mokri B., and Kurtzke J.: The epidemiology of Creutzfeldt-Jakob disease. *Slow Transmissible Diseases of the Nervous System, Vol. 1.* Prusiner S.B. and Hadlow W.J., Eds. Academic Press, New York, 1979 pp. 93–112.
10. Brown P., Cathala F., Raubertas R.F., Gajdusek D.C., and Castaigne P.: The epidemiology of Creutzfeldt-Jakob disease: conclusion of 15-year investigation in France and review of the world literature. *Neurology* 37:895–904, 1987.
11. Harries-Jones R., Knight R., Will R.G., Cousens S., Smith P.G., and Matthews W.B.: Creutzfeldt-Jakob disease in England and Wales, 1980–1984: a case-control study of potential risk factors. *J. Neurol. Neurosurg. Psychiatry* 51:1113–1119, 1988.
12. Cousens S.N., Harries-Jones R., Knight R., Will R.G., Smith P.G., and Matthews W.B.: Geographical distribution of cases of Creutzfeldt-Jakob disease in England and Wales 1970–84. *J. Neurol. Neurosurg. Psychiatry* 53:459–465, 1990.
13. Kahana E., Milton A., Braham J., and Sofer D.: Creutzfeldt-Jakob disease: focus among Libyan Jews in Israel. *Science* 183:90–91, 1974.
14. Goldfarb L., Korczyn A., Brown P., Chapman J., and Gajdusek D.C.: Mutation in codon 200 of scrapie amyloid precursor gene linked to Creutzfeldt-Jakob disease in Sephardic Jews of Libyan and non-Libyan origin. *Lancet* 336:637–638, 1990.
15. Hsiao K., Meiner Z., Kahana E., Cass C., Kahana I., Avrahami D., Scarlato G., Abramsky O., Prusiner S.B., and Gabizon R.: Mutation of the prion protein in Libyan Jews with Creutzfeldt-Jakob disease. *N. Engl. J. Med.* 324:1091–1097, 1991.
16. Gabizon R., Rosenmann H., Meiner Z., Kahana I., Kahana E., Shugart Y., Ott J., and Prusiner S.B.: Mutation and polymorphism of the prion protein gene in Libyan Jews with Creutzfeldt-Jakob disease. *Am. J. Hum. Genet.* 53:828–835, 1993.

17. Sigurdsson B.: Rida, a chronic encephalitis of sheep with general remarks on infections which develop slowly and some of their special characteristics. *Br. Vet. J.* 110:341–354, 1954.
18. Hadlow W.J.: Scrapie and kuru. *Lancet* 2:289–290, 1959.
19. Gajdusek D.C., Gibbs C.J., Jr., and Alpers M.: Experimental transmission of a kuru-like syndrome to chimpanzees. *Nature* 209:794–796, 1966.
20. Klatzo I., Gajdusek D.C., and Zigas V.: Pathology of kuru. *Lab. Invest.* 8:799–847, 1959.
21. Zlotnik I. and Stamp J.L.: Scrapie disease of sheep. *World Neurology* 2:895–907, 1961.
22. Masters C.L. and Richardson E.P., Jr.: Subacute spongiform encephalopathy Creutzfeldt-Jakob disease–the nature and progression of spongiform change. *Brain* 101:333–344, 1978.
23. Prusiner S.B.: Molecular biology of prion diseases. *Science* 252:1515–1522, 1991.
24. Wells G.A.H., Scott A.C., Johnson C.T., Gunning R.F., Hancock R.D., Jeffrey M., Dawson M., and Bradley R.: A novel progressive spongiform encephalopathy in cattle. *Vet. Rec.* 121:419–420, 1987.
25. Pan K.-M., Baldwin M., Nguyen J., Gasset M., Serban A., Groth D., Mehlhorn I., Huang Z., Fletterick R.J., Cohen F.E., and Prusiner S.B.: Conversion of α-helices into β-sheets features in the formation of the scrapie prion proteins. *Proc. Natl. Acad. Sci. USA* 90:10962–10966, 1993.
26. Bessen R.A. and Marsh R.F.: Distinct PrP properties suggest the molecular basis of strain variation in transmissible mink encephalopathy. *J. Virol.* 68:7859–7868, 1994.
27. Telling G.C., Parchi P., DeArmond S.J., Cortelli P., Montagna P., Gabizon R., Mastrianni J., Lugaresi E., Gambetti P., and Prusiner S.B.: Evidence for the conformation of the pathologic isoform of the prion protein enciphering and propagating prion diversity. *Science* 274:2079–2082, 1996.
28. Prusiner S.B.: Prion diseases and the BSE crisis. *Science* 278:245–251, 1997.
29. Dlouhy S.R., Hsiao K., Farlow M.R., Foroud T., Conneally P.M., Johnson P., Prusiner S.B., Hodes M.E., and Ghetti B.: Linkage of the Indiana kindred of Gerstmann-Sträussler-Scheinker disease to the prion protein gene. *Nat. Genet.* 1:64–67, 1992.
30. Petersen R.B., Tabaton M., Berg L., Schrank B., Torack R.M., Leal S., Julien J., Vital C., Deleplanque B., Pendlebury W.W., Drachman D., Smith T.W., Martin J.J., Oda M., Montagna P., Ott J., Autilio-Gambetti L., Lugaresi E., and Gambetti P.: Analysis of the prion protein gene in thalamic dementia. *Neurology* 42:1859–1863, 1992.
31. Poulter M., Baker H.F., Frith C.D., Leach M., Lofthouse R., Ridley R.M., Shah T., Owen F., Collinge J., Brown G., Hardy J., Mullan M.J., Harding A.E., Bennett C., Doshi R., and Crow T.J.: Inherited prion disease with 144 base pair gene insertion. 1. Genealogical and molecular studies. *Brain* 115:675–685, 1992.
32. Alper T., Haig D.A., and Clarke M.C.: The exceptionally small size of the scrapie agent. *Biochem. Biophys. Res. Commun.* 22:278–284, 1966.
33. Alper T., Cramp W.A., Haig D.A., and Clarke M.C.: Does the agent of scrapie replicate without nucleic acid? *Nature* 214:764–766, 1967.
34. Latarjet R., Muel B., Haig D.A., Clarke M.C., and Alper T.: Inactivation of the scrapie agent by near monochromatic ultraviolet light. *Nature* 227:1341–1343, 1970.
35. Gibbs C.J., Jr., Gajdusek D.C., and Latarjet R.: Unusual resistance to ionizing radiation of the viruses of kuru, Creutzfeldt-Jakob diseas. *Proc. Natl. Acad. Sci. USA* 75:6268–6270, 1978.
36. Pattison I.H.: Experiments with scrapie with special reference to the nature of the agent and the pathology of the disease. *Slow, Latent and Temperate Virus Infections, NINDB Monograph 2.* Gajdusek D.C., Gibbs C.J., Jr., and Alpers M.P., Eds. U.S. Government Printing, Washington, D.C., 1965 pp. 249–257.
37. Griffith J.S.: Self-replication and scrapie. *Nature* 215:1043–1044, 1967.
38. Pattison I.H. and Jones K.M.: The possible nature of the transmissible agent of scrapie. *Vet. Res.* 80:1–8, 1967.
39. Gibbons R.A. and Hunter G.D.: Nature of the scrapie agent. *Nature* 215:1041–1043, 1967.

40. Hunter G.D., Kimberlin R.H., and Gibbons R.A.: Scrapie: a modified membrane hypothesis. *J. Theor. Biol.* 20:355–357, 1968.
41. Field E.J., Farmer F., Caspary E.A., and Joyce G.: Susceptibility of scrapie agent to ionizing radiation. *Nature* 222:90–91, 1969.
42. Hunter G.D.: Scrapie: a prototype slow infection. *J. Infect. Dis.* 125:427–440, 1972.
43. Gordon W.S.: Advances in veterinary research. *Vet. Res.* 58:516–520, 1946.
44. Parry H.B.: Scrapie: a transmissible and hereditary disease of sheep. *Heredity* 17:75–105, 1962.
45. Dickinson A.G., Young G.B., Stamp J.T., and Renwick C.C.: An analysis of natural scrapie in Suffolk sheep. *Heredity* 20:485–503, 1965.
46. Parry H.B.: Recorded occurrences of scrapie from 1750. *Scrapie Disease in Sheep.* Oppenheimer D.R., Ed. Academic Press, New York, 1983 pp. 31–59.
47. Prusiner S.B.: An approach to the isolation of biological particles using sedimentation analysis. *J. Biol. Chem.* 253:916–921, 1978.
48. Prusiner S.B., Garfin D.E., Baringer J.R., Cochran S.P., Hadlow W.J., Race R.E., and Eklund C.M.: Evidence for multiple molecular forms of the scrapie agent. *Persistent Viruses.* Stevens J., Todaro G., and Fox C.F., Eds. Academic Press, New York, 1978 pp. 591–613.
49. Prusiner S.B.: Prions. *Sci. Am.* 251:50–59, 1984.
50. Chandler R.L.: Experimental scrapie in the mouse. *Res. Vet. Sci.* 4:276–285, 1963.
51. Prusiner S., Davis J.N., and Stadtman E.R.: Regulation of glutaminase B in *Escherichia coli.* I. Purification, properties, and cold liability. *J. Biol. Chem.* 251:3447–3456, 1976.
52. Prusiner S.B., Hadlow W.J., Eklund C.M., and Race R.E.: Sedimentation properties of the scrapie agent. *Proc. Natl. Acad. Sci. USA* 74:4656–4660, 1977.
53. Marsh R.F. and Kimberlin R.H.: Comparison of scrapie and transmissible mink encephalopathy in hamsters. II. Clinical signs, pathology and pathogenesis. *J. Infect. Dis.* 131:104–110, 1975.
54. Kimberlin R. and Walker C.: Characteristics of a short incubation model of scrapie in the golden hamster. *J. Gen. Virol.* 34:295–304, 1977.
55. Prusiner S.B., Groth D.F., Cochran S.P., Masiarz F.R., McKinley M.P., and Martinez H.M.: Molecular properties, partial purification, and assay by incubation period measurements of the hamster scrapie agent. *Biochemistry* 19:4883–4891, 1980.
56. Prusiner S.B., Cochran S.P., Groth D.F., Downey D.E., Bowman K.A., and Martinez H.M.: Measurement of the scrapie agent using an incubation time interval assay. *Ann. Neurol.* 11:353–358, 1982.
57. Zlotnik I.: Experimental transmission of scrapie to golden hamsters. *Lancet* 2:1072, 1963.
58. Prusiner S.B.: Novel proteinaceous infectious particles cause scrapie. *Science* 216:136–144, 1982.
59. Stahl N., Baldwin M.A., Teplow D.B., Hood L., Gibson B.W., Burlingame A.L., and Prusiner S.B.: Structural analysis of the scrapie prion protein using mass spectrometry and amino acid sequencing. *Biochemistry* 32:1991–2002, 1993.
60. Cohen F.E., Pan K.-M., Huang Z., Baldwin M., Fletterick R.J., and Prusiner S.B.: Structural clues to prion replication. *Science* 264:530–531, 1994.
61. Prusiner S.B., Groth D., Serban A., Stahl N., and Gabizon R.: Attempts to restore scrapie prion infectivity after exposure to protein denaturants. *Proc. Natl. Acad. Sci. USA* 90:2793–2797, 1993.
62. Carp R.I., Kascsak R.J., Rubenstein R., and Merz P.A.: The puzzle of PrP^{Sc} and infectivity–do the pieces fit? *Trends Neurosci.* 17:148–149, 1994.
63. Manuelidis L. and Fritch W.: Infectivity and host responses in Creutzfeldt-Jakob disease. *Virology* 216:46–59, 1996.
64. Narang H.: The nature of the scrapie agent: the virus theory. *Proc. Soc. Exp. Biol. Med.* 212:208–224, 1996.
65. Özel M., Baldauf E., Beekes M., and Diringer H.: Small virus-like particles in transmissible spongiform encephalopathies. *Transmissible Subacute Spongiform*

Encephalopathies: Prion Diseases. Court L. and Dodet B., Eds. Elsevier, Paris, 1996 pp. 369–373.

66. Caughey B. and Chesebro B.: Prion protein and the transmissible spongiform encephalopathies. *Trends Cell Biol.* 7:56–62, 1997.

67. Lasmézas C.I., Deslys J.-P., Robain O., Jaegly A., Beringue V., Peyrin J.-M., Fournier J.-G., Hauw J.-J., Rossier J., and Dormont D.: Transmission of the BSE agent to mice in the absence of detectable abnormal prion protein. *Science* 275:402–405, 1997.

68. Diener T.O., McKinley M.P., and Prusiner S.B.: Viroids and prions. *Proc. Natl. Acad. Sci. USA* 79:5220–5224, 1982.

69. Chamberlin T.C.: The method of multiple working hypotheses. *Science [Old Series]* 15:92–97, 1890.

70. Avery O.T., MacLeod C.M., and McCarty M.: Studies on the chemical nature of the substance inducing transformation of pneumococcal types. Induction of transformation by a deoxyribonucleic acid fraction isolated from pneumococcus type III. *J. Exp. Med.* 79:137–157, 1944.

71. Stanley W.M.: The "undiscovered" discovery. *Arch. Environ. Health* 21:256–262, 1970.

72. McCarty M.: *The Transforming Principle: Discovering that Genes Are Made of DNA,* W. W. Norton & Co., New York, 1985.

73. Hershey A.D. and Chase M.: Independent functions of viral protein and nucleic acid in growth of bacteriophage. *J. Gen. Physiol.* 36:39–56, 1952.

74. Watson J.D. and Crick F.H.C.: Genetical implication of the structure of deoxyribose nucleic acid. *Nature* 171:964–967, 1953.

75. Mirsky A.E. and Pollister A.W.: Chromosin, a desoxyribose nucleoprotein complex of the cell nucleus. *J. Gen. Physiol.* 30:134–135, 1946.

76. Diener T.O.: *Viroids and Viroid Diseases,* John Wiley, New York, 1979.

77. Fraenkel-Conrat H. and Williams R.C.: Reconstitution of active tobacco virus from the inactive protein and nucleic acid components. *Proc. Natl. Acad. Sci. USA* 41:690–698, 1955.

78. Gierer A. and Schramm G.: Infectivity of ribonucleic acid from tobacco mosaic virus. *Nature* 177:702–703, 1956.

79. Bolton D.C., McKinley M.P., and Prusiner S.B.: Identification of a protein that purifies with the scrapie prion. *Science* 218:1309–1311, 1982.

80. Prusiner S.B., Bolton D.C., Groth D.F., Bowman K.A., Cochran S.P., and McKinley M.P.: Further purification and characterization of scrapie prions. *Biochemistry* 21:6942–6950, 1982.

81. Prusiner S.B., Groth D.F., Bolton D.C., Kent S.B., and Hood L.E.: Purification and structural studies of a major scrapie prion protein. *Cell* 38:127–134, 1984.

82. Basler K., Oesch B., Scott M., Westaway D., Wälchli M., Groth D.F., McKinley M.P., Prusiner S.B., and Weissmann C.: Scrapie and cellular PrP isoforms are encoded by the same chromosomal gene. *Cell* 46:417–428, 1986.

83. Chesebro B., Race R., Wehrly K., Nishio J., Bloom M., Lechner D., Bergstrom S., Robbins K., Mayer L., Keith J.M., Garon C., and Haase A.: Identification of scrapie prion protein-specific mRNA in scrapie-infected and uninfected brain. *Nature* 315:331–333, 1985.

84. Oesch B., Westaway D., Wälchli M., McKinley M.P., Kent S.B.H., Aebersold R., Barry R.A., Tempst P., Teplow D.B., Hood L.E., Prusiner S.B., and Weissmann C.: A cellular gene encodes scrapie PrP 27–30 protein. *Cell* 40:735–746, 1985.

85. Meyer R.K., McKinley M.P., Bowman K.A., Braunfeld M.B., Barry R.A., and Prusiner S.B.: Separation and properties of cellular and scrapie prion proteins. *Proc. Natl. Acad. Sci. USA* 83:2310–2314, 1986.

86. Locht C., Chesebro B., Race R., and Keith J.M.: Molecular cloning and complete sequence of prion protein cDNA from mouse brain infected with the scrapie agent. *Proc. Natl. Acad. Sci. USA* 83:6372–6376, 1986.

87. Bendheim P.E., Barry R.A., DeArmond S.J., Stites D.P., and Prusiner S.B.: Antibodies to a scrapie prion protein. *Nature* 310:418–421, 1984.

88. Bockman J.M., Kingsbury D.T., McKinley M.P., Bendheim P.E., and Prusiner S.B.: Creutzfeldt-Jakob disease prion proteins in human brains. *N. Engl. J. Med.* 312:73–78, 1985.

89. Brown P., Coker-Vann M., Pomeroy K., Franko M., Asher D.M., Gibbs C.J., Jr., and Gajdusek D.C.: Diagnosis of Creutzfeldt-Jakob disease by Western blot identification of marker protein in human brain tissue. *N. Engl. J. Med.* 314:547–551, 1986.

90. Bockman J.M., Prusiner S.B., Tateishi J., and Kingsbury D.T.: Immunoblotting of Creutzfeldt-Jakob disease prion proteins: host species-specific epitopes. *Ann. Neurol.* 21:589–595, 1987.

91. Manuelidis L., Valley S., and Manuelidis E.E.: Specific proteins associated with Creutzfeldt-Jakob disease and scrapie share antigenic and carbohydrate determinants. *Proc. Natl. Acad. Sci. USA* 82:4263–4267, 1985.

92. Prusiner S.B.: Prions and neurodegenerative diseases. *N. Engl. J. Med.* 317:1571–1581, 1987.

93. Manuelidis L., Tesin D.M., Sklaviadis T., and Manuelidis E.E.: Astrocyte gene expression in Creutzfeldt-Jakob disease. *Proc. Natl. Acad. Sci. USA* 84:5937–5941, 1987.

94. Gomi H., Yokoyama T., Fujimoto K., Ikeda T., Katoh A., Itoh T., and Itohara S.: Mice devoid of the glial fibrillary acidic protein develop normally and are susceptible to scrapie prions. *Neuron* 14:29–41, 1995.

95. Tatzelt J., Maeda N., Pekny M., Yang S.-L., Betsholtz C., Eliasson C., Cayetano J., Camerino A.P., DeArmond S.J., and Prusiner S.B.: Scrapie in mice deficient in apolipoprotein E or glial fibrillary acidic protein. *Neurology* 47:449–453, 1996.

96. Duguid J.R., Rohwer R.G., and Seed B.: Isolation of cDNAs of scrapie-modulated RNAs by subtractive hybridization of a cDNA library. *Proc. Natl. Acad. Sci. USA* 85:5738–5742, 1988.

97. Duguid J.R., Bohmont C.W., Liu N., and Tourtellotte W.W.: Changes in brain gene expression shared by scrapie and Alzheimer disease. *Proc. Natl. Acad. Sci. USA* 86:7260–7264, 1989.

98. Diedrich J.F., Carp R.I., and Haase A.T.: Increased expression of heat shock protein, transferrin, and b$_2$-microglobulin in astrocytes during scrapie. *Microbial. Pathogenesis* 15:1–6, 1993.

99. Akowitz A., Sklaviadis T., and Manuelidis L.: Endogenous viral complexes with long RNA cosediment with the agent of Creutzfeldt-Jakob disease. *Nucleic Acids Res.* 22:1101–1107, 1994.

100. Büeler H., Aguzzi A., Sailer A., Greiner R.-A., Autenried P., Aguet M., and Weissmann C.: Mice devoid of PrP are resistant to scrapie. *Cell* 73:1339–1347, 1993.

101. Prusiner S.B., Groth D., Serban A., Koehler R., Foster D., Torchia M., Burton D., Yang S.-L., and DeArmond S.J.: Ablation of the prion protein (PrP) gene in mice prevents scrapie and facilitates production of anti-PrP antibodies. *Proc. Natl. Acad. Sci. USA* 90:10608–10612, 1993.

102. Manson J.C., Clarke A.R., McBride P.A., McConnell I., and Hope J.: PrP gene dosage determines the timing but not the final intensity or distribution of lesions in scrapie pathology. *Neurodegeneration* 3:331–340, 1994.

103. Sailer A., Büeler H., Fischer M., Aguzzi A., and Weissmann C.: No propagation of prions in mice devoid of PrP. *Cell* 77:967–968, 1994.

104. Sakaguchi S., Katamine S., Shigematsu K., Nakatani A., Moriuchi R., Nishida N., Kurokawa K., Nakaoke R., Sato H., Jishage K., Kuno J., Noda T., and Miyamoto T.: Accumulation of proteinase K-resistant prion protein (PrP) is restricted by the expression level of normal PrP in mice inoculated with a mouse-adapted strain of the Creutzfeldt–Jakob disease agent. *J. Virol.* 69:7586–7592, 1995.

105. Gabizon R., McKinley M.P., Groth D.F., and Prusiner S.B.: Immunoaffinity purification and neutralization of scrapie prion infectivity. *Proc. Natl. Acad. Sci. USA* 85:6617–6621, 1988.

106. McKinley M.P., Bolton D.C., and Prusiner S.B.: A protease-resistant protein is a structural component of the scrapie prion. *Cell* 35:57–62, 1983.

107. Prusiner S.B., McKinley M.P., Bowman K.A., Bolton D.C., Bendheim P.E., Groth D.F., and Glenner G.G.: Scrapie prions aggregate to form amyloid-like birefringent rods. *Cell* 35:349–358, 1983.

108. Bolton D.C., McKinley M.P., and Prusiner S.B.: Molecular characteristics of the major scrapie prion protein. *Biochemistry* 23:5898–5906, 1984.

109. Riesner D., Kellings K., Post K., Wille H., Serban H., Groth D., Baldwin M.A., and Prusiner S.B.: Disruption of prion rods generates 10-nm spherical particles having high α-helical content and lacking scrapie infectivity. *J. Virol.* 70:1714–1722, 1996.

110. Bellinger-Kawahara C., Cleaver J.E., Diener T.O., and Prusiner S.B.: Purified scrapie prions resist inactivation by UV irradiation. *J. Virol.* 61:159–166, 1987.

111. Bellinger-Kawahara C., Diener T.O., McKinley M.P., Groth D.F., Smith D.R., and Prusiner S.B.: Purified scrapie prions resist inactivation by procedures that hydrolyze, modify, or shear nucleic acids. *Virology* 160:271–274, 1987.

112. Bellinger-Kawahara C.G., Kempner E., Groth D.F., Gabizon R., and Prusiner S.B.: Scrapie prion liposomes and rods exhibit target sizes of 55,000 Da. *Virology* 164:537–541, 1988.

113. Kellings K., Meyer N., Mirenda C., Prusiner S.B., and Riesner D.: Further analysis of nucleic acids in purified scrapie prion preparations by improved return refocussing gel electrophoresis (RRGE). *J. Gen. Virol.* 73:1025–1029, 1992.

114. Kellings K., Prusiner S.B., and Riesner D.: Nucleic acids in prion preparations: unspecific background or essential component? *Phil. Trans. R. Soc. Lond.* B 343:425–430, 1994.

115. Manuelidis L., Sklaviadis T., Akowitz A., and Fritch W.: Viral particles are required for infection in neurodegenerative Creutzfeldt-Jakob disease. *Proc. Natl. Acad. Sci. USA* 92:5124–5128, 1995.

116. Williams R.C.: Electron microscopy of viruses. *Adv. Virus Res.* 2:183–239, 1954.

117. Cohen A.S., Shirahama T., and Skinner M.: Electron microscopy of amyloid. *Electron Microscopy of Proteins, Vol. 3.* Harris J.R., Ed. Academic Press, New York, 1982 pp. 165–206.

118. DeArmond S.J., McKinley M.P., Barry R.A., Braunfeld M.B., McColloch J.R., and Prusiner S.B.: Identification of prion amyloid filaments in scrapie-infected brain. *Cell* 41:221–235, 1985.

119. Kitamoto T., Tateishi J., Tashima I., Takeshita I., Barry R.A., DeArmond S.J., and Prusiner S.B.: Amyloid plaques in Creutzfeldt-Jakob disease stain with prion protein antibodies. *Ann. Neurol.* 20:204–208, 1986.

120. Roberts G.W., Lofthouse R., Brown R., Crow T.J., Barry R.A., and Prusiner S.B.: Prion-protein immunoreactivity in human transmissible dementias. *N. Engl. J. Med.* 315:1231–1233, 1986.

121. Gabizon R., McKinley M.P., and Prusiner S.B.: Purified prion proteins and scrapie infectivity copartition into liposomes. *Proc. Natl. Acad. Sci. USA* 84:4017–4021, 1987.

122. McKinley M.P., Meyer R., Kenaga L., Rahbar F., Cotter R., Serban A., and Prusiner S.B.: Scrapie prion rod formation *in vitro* requires both detergent extraction and limited proteolysis. *J. Virol.* 65:1440–1449, 1991.

123. Diringer H., Gelderblom H., Hilmert H., Ozel M., Edelbluth C., and Kimberlin R.H.: Scrapie infectivity, fibrils and low molecular weight protein. *Nature* 306:476–478, 1983.

124. Merz P.A., Kascsak R.J., Rubenstein R., Carp R.I., and Wisniewski H.M.: Antisera to scrapie-associated fibril protein and prion protein decorate scrapie-associated fibrils. *J. Virol.* 61:42–49, 1987.

125. Diener T.O.: PrP and the nature of the scrapie agent. *Cell* 49:719–721, 1987.

126. Merz P.A., Somerville R.A., Wisniewski H.M., and Iqbal K.: Abnormal fibrils from scrapie-infected brain. *Acta Neuropathol. (Berl.)* 54:63–74, 1981.

127. Merz P.A., Wisniewski H.M., Somerville R.A., Bobin S.A., Masters C.L., and Iqbal K.: Ultrastructural morphology of amyloid fibrils from neuritic and amyloid plaques. *Acta Neuropathol. (Berl.)* 60:113–124, 1983.

128. Merz P.A., Rohwer R.G., Kascsak R., Wisniewski H.M., Somerville R.A., Gibbs C.J., Jr.,

and Gajdusek D.C.: Infection-specific particle from the unconventional slow virus diseases. *Science* 225:437–440, 1984.

129. Diringer H.: Transmissible spongiform encephalopathies (TSE) virus-induced amyloidoses of the central nervous system (CNS). *Eur. J. Epidemiol.* 7:562–566, 1991.

130. Bastian F.O.: Bovine spongiform encephalopathy: relationship to human disease and nature of the agent. *ASM News* 59:235–240, 1993.

131. Kimberlin R.H.: Scrapie and possible relationships with viroids. *Semin. Virol.* 1:153–162, 1990.

132. Chesebro B. and Caughey B.: Scrapie agent replication without the prion protein? *Curr. Biol.* 3:696–698, 1993.

133. Scott M., Groth D., Foster D., Torchia M., Yang S.-L., DeArmond S.J., and Prusiner S.B.: Propagation of prions with artificial properties in transgenic mice expressing chimeric PrP genes. *Cell* 73:979–988, 1993.

134. Telling G.C., Scott M., Mastrianni J., Gabizon R., Torchia M., Cohen F.E., DeArmond S.J., and Prusiner S.B.: Prion propagation in mice expressing human and chimeric PrP transgenes implicates the interaction of cellular PrP with another protein. *Cell* 83:79–90, 1995.

135. Scott M.R., Groth D., Tatzelt J., Torchia M., Tremblay P., DeArmond S.J., and Prusiner S.B.: Propagation of prion strains through specific conformers of the prion protein. *J. Virol.* 71:9032–9044, 1997.

136. Anfinsen C.B.: Principles that govern the folding of protein chains. *Science* 181:223–230, 1973.

137. Pergami P., Jaffe H., and Safar J.: Semipreparative chromatographic method to purify the normal cellular isoform of the prion protein in nondenatured form. *Anal. Biochem.* 236:63–73, 1996.

138. Borchelt D.R., Scott M., Taraboulos A., Stahl N., and Prusiner S.B.: Scrapie and cellular prion proteins differ in their kinetics of synthesis and topology in cultured cells. *J. Cell Biol.* 110:743–752, 1990.

139. Stahl N., Borchelt D.R., Hsiao K., and Prusiner S.B.: Scrapie prion protein contains a phosphatidylinositol glycolipid. *Cell* 51:229–240, 1987.

140. Caughey B. and Raymond G.J.: The scrapie-associated form of PrP is made from a cell surface precursor that is both protease- and phospholipase-sensitive. *J. Biol. Chem.* 266:18217–18223, 1991.

141. Borchelt D.R., Taraboulos A., and Prusiner S.B.: Evidence for synthesis of scrapie prion proteins in the endocytic pathway. *J. Biol. Chem.* 267:16188–16199, 1992.

142. Taraboulos A., Scott M., Semenov A., Avrahami D., Laszlo L., and Prusiner S.B.: Cholesterol depletion and modification of COOH-terminal targeting sequence of the prion protein inhibits formation of the scrapie isoform. *J. Cell Biol.* 129:121–132, 1995.

143. Vey M., Pilkuhn S., Wille H., Nixon R., DeArmond S.J., Smart E.J., Anderson R.G., Taraboulos A., and Prusiner S.B.: Subcellular colocalization of the cellular and scrapie prion proteins in caveolae-like membranous domains. *Proc. Natl. Acad. Sci. USA* 93:14945–14949, 1996.

144. Kaneko K., Vey M., Scott M., Pilkuhn S., Cohen F.E., and Prusiner S.B.: COOH-terminal sequence of the cellular prion protein directs subcellular trafficking and controls conversion into the scrapie isoform. *Proc. Natl. Acad. Sci. USA* 94:2333–2338, 1997.

145. Naslavsky N., Stein R., Yanai A., Friedlander G., and Taraboulos A.: Characterization of detergent-insoluble complexes containing the cellular prion protein and its scrapie isoform. *J. Biol. Chem.* 272:6324–6331, 1997.

146. Gasset M., Baldwin M.A., Lloyd D., Gabriel J.-M., Holtzman D.M., Cohen F., Fletterick R., and Prusiner S.B.: Predicted α-helical regions of the prion protein when synthesized as peptides form amyloid. *Proc. Natl. Acad. Sci. USA* 89:10940–10944, 1992.

147. Huang Z., Gabriel J.-M., Baldwin M.A., Fletterick R.J., Prusiner S.B., and Cohen F.E.: Proposed three-dimensional structure for the cellular prion protein. *Proc. Natl. Acad. Sci. USA* 91:7139–7143, 1994.

148. Zhang H., Kaneko K., Nguyen J.T., Livshits T.L., Baldwin M.A., Cohen F.E., James T.L.,

and Prusiner S.B.: Conformational transitions in peptides containing two putative α-helices of the prion protein. *J. Mol. Biol.* 250:514–526, 1995.

149. Schätzl H.M., Da Costa M., Taylor L., Cohen F.E., and Prusiner S.B.: Prion protein gene variation among primates. *J. Mol. Biol.* 245:362–374, 1995.

150. Bamborough P., Wille H., Telling G.C., Yehiely F., Prusiner S.B., and Cohen F.E.: Prion protein structure and scrapie replication: theoretical, spectroscopic and genetic investigations. *Cold Spring Harb. Symp. Quant. Biol.* 61:495–509, 1996.

151. Hay B., Barry R.A., Lieberburg I., Prusiner S.B., and Lingappa V.R.: Biogenesis and transmembrane orientation of the cellular isoform of the scrapie prion protein. *Mol. Cell. Biol.* 7:914–920, 1987.

152. De Fea K.A., Nakahara D.H., Calayag M.C., Yost C.S., Mirels L.F., Prusiner S.B., and Lingappa V.R.: Determinants of carboxyl-terminal domain translocation during prion protein biogenesis. *J. Biol. Chem.* 269:16810–16820, 1994.

153. Hegde R.S., Mastrianni J.A., Scott M.R., DeFea K.A., Tremblay P., Torchia M., DeArmond S.J., Prusiner S.B., and Lingappa V.R.: A transmembrane form of the prion protein in neurodegenerative disease. *Science* 279: 827–834, 1998.

154. Riek R., Hornemann S., Wider G., Billeter M., Glockshuber R., and Wüthrich K.: NMR structure of the mouse prion protein domain PrP(121–231). *Nature* 382:180–182, 1996.

155. James T.L., Liu H., Ulyanov N.B., Farr-Jones S., Zhang H., Donne D.G., Kaneko K., Groth D., Mehlhorn I., Prusiner S.B., and Cohen F.E.: Solution structure of a 142-residue recombinant prion protein corresponding to the infectious fragment of the scrapie isoform. *Proc. Natl. Acad. Sci. USA* 94:10086–10091, 1997.

156. Donne D.G., Viles J.H., Groth D., Mehlhorn I., James T.L., Cohen F.E., Prusiner S.B., Wright P.E., and Dyson H.J.: Structure of the recombinant full-length hamster prion protein PrP(29–231): the N terminus is highly flexible. *Proc. Natl. Acad. Sci. USA* 94:13452–13457, 1997.

157. Williamson R.A., Peretz D., Smorodinsky N., Bastidas R., Serban H., Mehlhorn I., DeArmond S.J., Prusiner S.B., and Burton D.R.: Circumventing tolerance to generate autologous monoclonal antibodies to the prion protein. *Proc. Natl. Acad. Sci. USA* 93:7279–7282, 1996.

158. Peretz D., Williamson R.A., Matsunaga Y., Serban H., Pinilla C., Bastidas R., Rozenshteyn R., James T.L., Houghten R.A., Cohen F.E., Prusiner S.B., and Burton D.R.: A conformational transition at the N terminus of the prion protein features in formation of the scrapie isoform. *J. Mol. Biol.* 273:614–622, 1997.

159. Caughey B., Race R.E., Ernst D., Buchmeier M.J., and Chesebro B.: Prion protein biosynthesis in scrapie-infected and uninfected neuroblastoma cells. *J. Virol.* 63:175–181, 1989.

160. Shmerling D., Hegyi I., Fischer M., Blättler T., Brandner S., Götz J., Rülicke T., Flechsig E., Cozzio A., von Mering C., Hangartner C., Aguzzi A., and Weissmann C.: Expression of amino-terminally truncated PrP in the mouse leading to ataxia and specific cerebellar lesions. *Cell* 93:203–214, 1998.

161. Mehlhorn I., Groth D., Stöckel J., Moffat B., Reilly D., Yansura D., Willett W.S., Baldwin M., Fletterick R., Cohen F.E., Vandlen R., Henner D., and Prusiner S.B.: High-level expression and characterization of a purified 142-residue polypeptide of the prion protein. *Biochemistry* 35:5528–5537, 1996.

162. Zhang H., Stöckel J., Mehlhorn I., Groth D., Baldwin M.A., Prusiner S.B., James T.L., and Cohen F.E.: Physical studies of conformational plasticity in a recombinant prion protein. *Biochemistry* 36:3543–3553, 1997.

163. Fischer M., Rülicke T., Raeber A., Sailer A., Moser M., Oesch B., Brandner S., Aguzzi A., and Weissmann C.: Prion protein (PrP) with amino-proximal deletions restoring susceptibility of PrP knockout mice to scrapie. *EMBO J.* 15:1255–1264, 1996.

164. Muramoto T., DeArmond S.J., Scott M., Telling G.C., Cohen F.E., and Prusiner S.B.: Heritable disorder resembling neuronal storage disease in mice expressing prion protein with deletion of an α-helix. *Nat. Med.* 3:750–755, 1997.

165. Riek R., Hornemann S., Wider G., Glockshuber R., and Wüthrich K.: NMR characterization of the full-length recombinant murine prion protein, *m*PrP(23–231). *FEBS Lett* 413:282–288, 1997.

166. Huang Z., Prusiner S.B., and Cohen F.E.: Scrapie prions: a three-dimensional model of an infectious fragment. *Folding & Design* 1:13–19, 1996.

167. Turk E., Teplow D.B., Hood L.E., and Prusiner S.B.: Purification and properties of the cellular and scrapie hamster prion proteins. *Eur. J. Biochem.* 176:21–30, 1988.

168. Muramoto T., Scott M., Cohen F., and Prusiner S.B.: Recombinant scrapie-like prion protein of 106 amino acids is soluble. *Proc. Natl. Acad. Sci. USA* 93:15457–15462, 1996.

169. Caughey B.W., Dong A., Bhat K.S., Ernst D., Hayes S.F., and Caughey W.S.: Secondary structure analysis of the scrapie-associated protein PrP 27–30 in water by infrared spectroscopy. *Biochemistry* 30:7672–7680, 1991.

170. Gasset M., Baldwin M.A., Fletterick R.J., and Prusiner S.B.: Perturbation of the secondary structure of the scrapie prion protein under conditions associated with changes in infectivity. *Proc. Natl. Acad. Sci. USA* 90:1–5, 1993.

171. Safar J., Roller P.P., Gajdusek D.C., and Gibbs C.J., Jr.: Conformational transitions, dissociation, and unfolding of scrapie amyloid (prion) protein. *J. Biol. Chem.* 268:20276–20284, 1993.

172. Rogers M., Yehiely F., Scott M., and Prusiner S.B.: Conversion of truncated and elongated prion proteins into the scrapie isoform in cultured cells. *Proc. Natl. Acad. Sci. USA* 90:3182–3186, 1993.

173. Barry R.A. and Prusiner S.B.: Monoclonal antibodies to the cellular and scrapie prion proteins. *J. Infect. Dis.* 154:518–521, 1986.

174. Kascsak R.J., Rubenstein R., Merz P.A., Tonna-DeMasi M., Fersko R., Carp R.I., Wisniewski H.M., and Diringer H.: Mouse polyclonal and monoclonal antibody to scrapie-associated fibril proteins. *J. Virol.* 61:3688–3693, 1987.

175. Rogers M., Serban D., Gyuris T., Scott M., Torchia T., and Prusiner S.B.: Epitope mapping of the Syrian hamster prion protein utilizing chimeric and mutant genes in a vaccinia virus expression system. *J. Immunol.* 147:3568–3574, 1991.

176. Korth C., Stierli B., Streit P., Moser M., Schaller O., Fischer R., Schulz-Schaeffer W., Kretzschmar H., Raeber A., Braun U., Ehrensperger F., Hornemann S., Glockshuber R., Riek R., Billeter M., Wuthrick K., and Oesch B.: Prion (PrP^{Sc})-specific epitope defined by a monoclonal antibody. *Nature* 389:74–77, 1997.

177. Billeter M., Riek R., Wider G., Hornemann S., Glockshuber R., and Wüthrich K.: Prion protein NMR structure and species barrier for prion diseases. *Proc. Natl. Acad. Sci. USA* 94:7281–7285, 1997.

178. Kaneko K., Zulianello L., Scott M., Cooper C.M., Wallace A.C., James T.L., Cohen F.E., and Prusiner S.B.: Evidence for protein X binding to a discontinuous epitope on the cellular prion protein during scrapie prion propagation. *Proc. Natl. Acad. Sci. USA* 94:10069–10074, 1997.

179. Telling G.C., Scott M., Hsiao K.K., Foster D., Yang S.-L., Torchia M., Sidle K.C.L., Collinge J., DeArmond S.J., and Prusiner S.B.: Transmission of Creutzfeldt-Jakob disease from humans to transgenic mice expressing chimeric human-mouse prion protein. *Proc. Natl. Acad. Sci. USA* 91:9936–9940, 1994.

180. Stöckel J., Safar J., Wallace A.C., Cohen F.E., and Prusiner S.B.: Prion protein selectively binds copper-II-ions. *Biochemistry* 37: 7185–7193, 1998.

181. Hornshaw M.P., McDermott J.R., Candy J.M., and Lakey J.H.: Copper binding to the N-terminal tandem repeat region of mammalian and avian prion protein: structural studies using synthetic peptides. *Biochem. Biophys. Res. Commun.* 214:993–999, 1995.

182. Hornshaw M.P., McDermott J.R., and Candy J.M.: Copper binding to the N-terminal tandem repeat regions of mammalian and avian prion protein. *Biochem. Biophys. Res. Commun.* 207:621–629, 1995.

183. Miura T., Hori-i A., and Takeuchi H.: Metal-dependent α-helix formation promoted by the glycine-rich octapeptide region of prion protein. *FEBS Lett.* 396:248–252, 1996.

184. Brown D.R., Schulz-Schaeffer W.J., Schmidt B., and Kretzschmar H.A.: Prion protein-

deficient cells show altered response to oxidative stress due to decreased SOD-1 activity. *Exp. Neurol.* 146:104–112, 1997.

185. Harris E.D.: Copper as a cofactor and regulator of copper, zinc superoxide dismutase. *J. Nutr.* 122:636–640, 1992.

186. Brown D.R., Qin K., Herms J.W., Madlung A., Manson J., Strome R., Fraser P.E., Kruck T., von Bohlen A., Schulz-Schaeffer W., Giese A., Westaway D., and Kretzschmar H.: The cellular prion protein binds copper *in vivo. Nature* 390:684–687, 1997.

187. Vulpe C., Levinson B., Whitney S., Packman S., and Gitschier J.: Isolation of a candidate gene for Menkes disease and evidence that it encodes a copper-transporting ATPase. *Nat. Genet.* 3:7–13, 1993.

188. Chelly J., Tümer Z., Tønnesen T., Petterson A., Ishikawa-Brush Y., Tommerup N., Horn N., and Monaco A.P.: Isolation of a candidate gene for Menkes disease that encodes a potential heavy metal binding protein. *Nat. Genet.* 3:14–19, 1993.

189. Bull P.C., Thomas G.R., Rommens J.M., Forbes J.R., and Cox D.W.: The Wilson disease gene is a putative copper transporting P-type ATPase similar to the Menkes gene. *Nat. Genet.* 5:327–337, 1993.

190. Petrukhin K., Fischer S.G., Pirastu M., Tanzi R.E., Chernov I., Devoto M., Brzustowicz L.M., Cayanis E., Vitale E., Russo J.J., Matseoane D., Boukhgalter B., Wasco W., Figus A.L., Loudianos J., Cao A., Sternlieb I., Evgrafov O., Parano E., Pavone L., Warburton D., Ott J., Penchaszadeh G.K., Scheinberg I.H., and Gilliam T.C.: Mapping, cloning and genetic characterization of the region containing the Wilson disease gene. *Nat. Genet.* 5:338–343, 1993.

191. Pattison I.H. and Jebbett J.N.: Clinical and histological observations on cuprizone toxicity and scrapie in mice. *Res. Vet. Sci.* 12:378–380, 1971.

192. Kimberlin R.H., Millson G.C., Bountiff L., and Collis S.C.: A comparison of the biochemical changes induced in mouse brain by cuprizone toxicity and by scrapie infection. *J. Comp. Path.* 84:263–270, 1974.

193. Gabriel J.-M., Oesch B., Kretzschmar H., Scott M., and Prusiner S.B.: Molecular cloning of a candidate chicken prion protein. *Proc. Natl. Acad. Sci. USA* 89:9097–9101, 1992.

194. Westaway D., Goodman P.A., Mirenda C.A., McKinley M.P., Carlson G.A., and Prusiner S.B.: Distinct prion proteins in short and long scrapie incubation period mice. *Cell* 51:651–662, 1987.

195. Westaway D., Mirenda C.A., Foster D., Zebarjadian Y., Scott M., Torchia M., Yang S.-L., Serban H., DeArmond S.J., Ebeling C., Prusiner S.B., and Carlson G.A.: Paradoxical shortening of scrapie incubation times by expression of prion protein transgenes derived from long incubation period mice. *Neuron* 7:59–68, 1991.

196. Yoshimoto J., Iinuma T., Ishiguro N., Horiuchi M., Imamura M., and Shinagawa M.: Comparative sequence analysis and expression of bovine PrP gene in mouse L-929 cells. *Virus Genes* 6:343–356, 1992.

197. Westaway D., Cooper C., Turner S., Da Costa M., Carlson G.A., and Prusiner S.B.: Structure and polymorphism of the mouse prion protein gene. *Proc. Natl. Acad. Sci. USA* 91:6418–6422, 1994.

198. Westaway D., Zuliani V., Cooper C.M., Da Costa M., Neuman S., Jenny A.L., Detwiler L., and Prusiner S.B.: Homozygosity for prion protein alleles encoding glutamine-171 renders sheep susceptible to natural scrapie. *Genes Dev.* 8:959–969, 1994.

199. Saeki K., Matsumoto Y., Hirota Y., Matsumoto Y., and Onodera T.: Three-exon structure of the gene encoding the rat prion protein and its expression in tissues. *Virus Genes* 12:15–20, 1996.

200. Li G. and Bolton D.C.: A novel hamster prion protein mRNA contains an extra exon: increased expression in scrapie. *Brain Res.* 751:265–274, 1997.

201. Lee I., Westaway D., Smit A., Wang K., Cooper C., Yao H., Prusiner S.B., and Hood L.: in preparation.

202. McKnight S. and Tjian R.: Transcriptional selectivity of viral genes in mammalian cells. *Cell* 46:795–805, 1986.

203. Robakis N.K., Devine-Gage E.A., Kascsak R.J., Brown W.T., Krawczun C., and Silverman W.P.: Localization of a human gene homologous to the PrP gene on the p arm of chromosome 20 and detection of PrP-related antigens in normal human brain. *Biochem. Biophys. Res. Commun.* 140:758–765, 1986.

204. Sparkes R.S., Simon M., Cohn V.H., Fournier R.E.K., Lem J., Klisak I., Heinzmann C., Blatt C., Lucero M., Mohandas T., DeArmond S.J., Westaway D., Prusiner S.B., and Weiner L.P.: Assignment of the human and mouse prion protein genes to homologous chromosomes. *Proc. Natl. Acad. Sci. USA* 83:7358–7362, 1986.

205. Mobley W.C., Neve R.L., Prusiner S.B., and McKinley M.P.: Nerve growth factor increases mRNA levels for the prion protein and the beta-amyloid protein precursor in developing hamster brain. *Proc. Natl. Acad. Sci. USA* 85:9811–9815, 1988.

206. Kretzschmar H.A., Prusiner S.B., Stowring L.E., and DeArmond S.J.: Scrapie prion proteins are synthesized in neurons. *Am. J. Pathol.* 122:1–5, 1986.

207. DeArmond S.J., Mobley W.C., DeMott D.L., Barry R.A., Beckstead J.H., and Prusiner S.B.: Changes in the localization of brain prion proteins during scrapie infection. *Neurology* 37:1271–1280, 1987.

208. Taraboulos A., Jendroska K., Serban D., Yang S.-L., DeArmond S.J., and Prusiner S.B.: Regional mapping of prion proteins in brains. *Proc. Natl. Acad. Sci. USA* 89:7620–7624, 1992.

209. Kimberlin R.H., Field H.J., and Walker C.A.: Pathogenesis of mouse scrapie: evidence for spread of infection from central to peripheral nervous system. *J. Gen. Virol.* 64:713–716, 1983.

210. Fraser H. and Dickinson A.G.: Targeting of scrapie lesions and spread of agent via the retino-tectal projection. *Brain Res.* 346:32–41, 1985.

211. Jendroska K., Heinzel F.P., Torchia M., Stowring L., Kretzschmar H.A., Kon A., Stern A., Prusiner S.B., and DeArmond S.J.: Proteinase-resistant prion protein accumulation in Syrian hamster brain correlates with regional pathology and scrapie infectivity. *Neurology* 41:1482–1490, 1991.

212. Carlson G.A., Kingsbury D.T., Goodman P.A., Coleman S., Marshall S.T., DeArmond S.J., Westaway D., and Prusiner S.B.: Linkage of prion protein and scrapie incubation time genes. *Cell* 46:503–511, 1986.

213. Scott M., Foster D., Mirenda C., Serban D., Coufal F., Wälchli M., Torchia M., Groth D., Carlson G., DeArmond S.J., Westaway D., and Prusiner S.B.: Transgenic mice expressing hamster prion protein produce species-specific scrapie infectivity and amyloid plaques. *Cell* 59:847–857, 1989.

214. Dickinson A.G., Meikle V.M.H., and Fraser H.: Identification of a gene which controls the incubation period of some strains of scrapie agent in mice. *J. Comp. Pathol.* 78:293–299, 1968.

215. Kingsbury D.T., Kasper K.C., Stites D.P., Watson J.D., Hogan R.N., and Prusiner S.B.: Genetic control of scrapie and Creutzfeldt-Jakob disease in mice. *J. Immunol.* 131:491–496, 1983.

216. Carlson G.A., Goodman P.A., Lovett M., Taylor B.A., Marshall S.T., Peterson-Torchia M., Westaway D., and Prusiner S.B.: Genetics and polymorphism of the mouse prion gene complex: the control of scrapie incubation time. *Mol. Cell. Biol.* 8:5528–5540, 1988.

217. Carlson G.A., Ebeling C., Yang S.-L., Telling G., Torchia M., Groth D., Westaway D., DeArmond S.J., and Prusiner S.B.: Prion isolate specified allotypic interactions between the cellular and scrapie prion proteins in congenic and transgenic mice. *Proc. Natl. Acad. Sci. USA* 91:5690–5694, 1994.

218. Moore R.C., Hope J., McBride P.A., McConnell I., Selfridge J., and Melton D.W.: Mice with gene targeted prion protein alterations show that *Prn-p*, *Sinc* and *Prn-i* are congruent. *Nat. Genet.* 18:118–125, 1998.

219. Prusiner S.B., Scott M., Foster D., Pan K.-M., Groth D., Mirenda C., Torchia M., Yang S.-L., Serban D., Carlson G.A., Hoppe P.C., Westaway D., and DeArmond S.J.: Transgenetic studies implicate interactions between homologous PrP isoforms in scrapie prion replication. *Cell* 63:673–686, 1990.

220. Büeler H., Fischer M., Lang Y., Bluethmann H., Lipp H.-P., DeArmond S.J., Prusiner S.B., Aguet M., and Weissmann C.: Normal development and behaviour of mice lacking the neuronal cell-surface PrP protein. *Nature* 356:577–582, 1992.

221. Manson J.C., Clarke A.R., Hooper M.L., Aitchison L., McConnell I., and Hope J.: 129/Ola mice carrying a null mutation in PrP that abolishes mRNA production are developmentally normal. *Mol. Neurobiol.* 8:121–127, 1994.

222. Sakaguchi S., Katamine S., Nishida N., Moriuchi R., Shigematsu K., Sugimoto T., Nakatani A., Kataoka Y., Houtani T., Shirabe S., Okada H., Hasegawa S., Miyamoto T., and Noda T.: Loss of cerebellar Purkinje cells in aged mice homozygous for a disrupted PrP gene. *Nature* 380:528–531, 1996.

223. Tobler I., Gaus S.E., Deboer T., Achermann P., Fischer M., Rülicke T., Moser M., Oesch B., McBride P.A., and Manson J.C.: Altered circadian activity rhythms and sleep in mice devoid of prion protein. *Nature* 380:639–642, 1996.

224. Collinge J., Whittington M.A., Sidle K.C., Smith C.J., Palmer M.S., Clarke A.R., and Jefferys J.G.R.: Prion protein is necessary for normal synaptic function. *Nature* 370:295–297, 1994.

225. Whittington M.A., Sidle K.C.L., Gowland I., Meads J., Hill A.F., Palmer M.S., Jefferys J.G.R., and Collinge J.: Rescue of neurophysiological phenotype seen in PrP null mice by transgene encoding human prion protein. *Nat. Genet.* 9:197–201, 1995.

226. Herms J.W., Kretzschmar H.A., Titz S., and Keller B.U.: Patch-clamp analysis of synaptic transmission to cerebellar purkinje cells of prion protein knockout mice. *Eur. J. Neurosci.* 7:2508–2512, 1995.

227. Lledo P.-M., Tremblay P., DeArmond S.J., Prusiner S.B., and Nicoll R.A.: Mice deficient for prion protein exhibit normal neuronal excitability and synaptic transmission in the hippocampus. *Proc. Natl. Acad. Sci. USA* 93:2403–2407, 1996.

228. Pattison I.H. and Jones K.M.: Modification of a strain of mouse-adapted scrapie by passage through rats. *Res. Vet. Sci.* 9:408–410, 1968.

229. Kimberlin R.H., Walker C.A., and Fraser H.: The genomic identity of different strains of mouse scrapie is expressed in hamsters and preserved on reisolation in mice. *J. Gen. Virol.* 70:2017–2025, 1989.

230. Hecker R., Taraboulos A., Scott M., Pan K.-M., Torchia M., Jendroska K., DeArmond S.J., and Prusiner S.B.: Replication of distinct prion isolates is region specific in brains of transgenic mice and hamsters. *Genes Dev.* 6:1213–1228, 1992.

231. Hunter N., Goldmann W., Benson G., Foster J.D., and Hope J.: Swaledale sheep affected by natural scrapie differ significantly in PrP genotype frequencies from healthy sheep and those selected for reduced incidence of scrapie. *J. Gen. Virol.* 74:1025–1031, 1993.

232. Shibuya S., Higuchi J., Shin R.-W., Tateishi J., and Kitamoto T.: Protective prion protein polymorphisms against sporadic Creutzfeldt-Jakob disease. *Lancet* 351:419, 1998.

233. Tatzelt J., Zuo J., Voellmy R., Scott M., Hartl U., Prusiner S.B., and Welch W.J.: Scrapie prions selectively modify the stress response in neuroblastoma cells. *Proc. Natl. Acad. Sci. USA* 92:2944–2948, 1995.

234. Tatzelt J., Prusiner S.B., and Welch W.J.: Chemical chaperones interfere with the formation of scrapie prion protein. *EMBO J.* 15:6363–6373, 1996.

235. Kenward N., Hope J., Landon M., and Mayer R.J.: Expression of polyubiquitin and heat-shock protein 70 genes increases in the later stages of disease progression in scrapie-infected mouse brain. *J. Neurochem.* 62:1870–1877, 1994.

236. Oesch B., Teplow D.B., Stahl N., Serban D., Hood L.E., and Prusiner S.B.: Identification of cellular proteins binding to the scrapie prion protein. *Biochemistry* 29:5848–5855, 1990.

237. Kurschner C. and Morgan J.I.: Analysis of interaction sites in homo- and heteromeric complexes containing Bcl-2 family members and the cellular prion protein. *Mol. Brain Res.* 37:249–258, 1996.

238. Edenhofer F., Rieger R., Famulok M., Wendler W., Weiss S., and Winnacker E.-L.: Prion protein PrP^C interacts with molecular chaperones of the Hsp60 family. *J. Virol.* 70:4724–4728, 1996.

239. Gerstmann J., Sträussler E., and Scheinker I.: Über eine eigenartige hereditär-familiäre Erkrankung des Zentralnervensystems zugleich ein Beitrag zur frage des vorzeitigen lokalen Alterns. *Z. Neurol.* 154:736–762, 1936.

240. Masters C.L., Harris J.O., Gajdusek D.C., Gibbs C.J., Jr., Bernouilli C., and Asher D.M.: Creutzfeldt-Jakob disease: patterns of worldwide occurrence and the significance of familal and sporadic clustering. *Ann. Neurol.* 5:177–188, 1978.

241. Gajdusek D.C.: Unconventional viruses and the origin and disappearance of kuru. *Science* 197:943–960, 1977.

242. Bobowick A.R., Brody J.A., Matthews M.R., Roos R., and Gajdusek D.C.: Creutzfeldt-Jakob disease: a case-control study. *Am. J. Epidemiol.* 98:381–394, 1973.

243. Rosenthal N.P., Keesey J., Crandall B., and Brown W.J.: Familial neurological disease associated with spongiform encephalopathy. *Arch. Neurol.* 33:252–259, 1976.

244. Doh-ura K., Tateishi J., Sasaki H., Kitamoto T., and Sakaki Y.: Pro->Leu change at position 102 of prion protein is the most common but not the sole mutation related to Gerstmann-Sträussler syndrome. *Biochem. Biophys. Res. Commun.* 163:974–979, 1989.

245. Goldgaber D., Goldfarb L.G., Brown P., Asher D.M., Brown W.T., Lin S., Teener J.W., Feinstone S.M., Rubenstein R., Kascsak R.J., Boellaard J.W., and Gajdusek D.C.: Mutations in familial Creutzfeldt-Jakob disease and Gerstmann-Sträussler-Scheinker's syndrome. *Exp. Neurol.* 106:204–206, 1989.

246. Kretzschmar H.A., Honold G., Seitelberger F., Feucht M., Wessely P., Mehraein P., and Budka H.: Prion protein mutation in family first reported by Gerstmann, Sträussler, and Scheinker. *Lancet* 337:1160, 1991.

247. Hsiao K.K., Scott M., Foster D., Groth D.F., DeArmond S.J., and Prusiner S.B.: Spontaneous neurodegeneration in transgenic mice with mutant prion protein. *Science* 250:1587–1590, 1990.

248. Hsiao K.K., Groth D., Scott M., Yang S.-L., Serban H., Rapp D., Foster D., Torchia M., DeArmond S.J., and Prusiner S.B.: Serial transmission in rodents of neurodegeneration from transgenic mice expressing mutant prion protein. *Proc. Natl. Acad. Sci. USA* 91:9126–9130, 1994.

249. Telling G.C., Haga T., Torchia M., Tremblay P., DeArmond S.J., and Prusiner S.B.: Interactions between wild-type and mutant prion proteins modulate neurodegeneration in transgenic mice. *Genes & Dev.* 10:1736–1750, 1996.

250. Scott M.R., Nguyen O., Stöckel J., Tatzelt J., DeArmond S.J., Cohen F.E., and Prusiner S.B.: Designer mutations in the prion protein promote β-sheet formation *in vitro* and cause neurodegeneration in transgenic mice (Abstr.). *Protein Sci.* 6 [Suppl. 1]:84, 1997.

251. Alpers M.P.: Kuru: implications of its transmissibility for interpretation of its changing epidemiological pattern. *The Central Nervous System, Some Experimental Models of Neurological Diseases.* Bailey O.T. and Smith D.E., Eds. Williams and Wilkins, Baltimore, 1968 pp. 234–251.

252. Alpers M.: Epidemiology and clinical aspects of kuru. *Prions–Novel Infectious Pathogens Causing Scrapie and Creutzfeldt-Jakob Disease.* Prusiner S.B. and McKinley M.P., Eds. Academic Press, Orlando, 1987 pp. 451–465.

253. Brown P., Preece M.A., and Will R.G.: "Friendly fire" in medicine: hormones, homografts, and Creutzfeldt-Jakob disease. *Lancet* 340:24–27, 1992.

254. Billette de Villemeur T., Deslys J.-P., Pradel A., Soubrié C., Alpérovitch A., Tardieu M., Chaussain J.-L., Hauw J.-J., Dormont D., Ruberg M., and Agid Y.: Creutzfeldt-Jakob disease from contaminated growth hormone extracts in France. *Neurology* 47:690–695, 1996.

255. Prusiner S. B., Scott M. R., Dearmond S. J., and Cohen F. E. Prion protein biology. *Cell* 93:337–348, 1998.

256. Esmonde T., Lueck C.J., Symon L., Duchen L.W., and Will R.G.: Creutzfeldt-Jakob dis-

ease and lyophilised dura mater grafts: report of two cases. *J. Neurol. Neurosurg. Psychiatry* 56:999–1000, 1993.

257. Lane K.L., Brown P., Howell D.N., Crain B.J., Hulette C.M., Burger P.C., and DeArmond S.J.: Creutzfeldt-Jakob disease in a pregnant woman with an implanted dura mater graft. *Neurosurgery* 34:737–740, 1994.

258. CDC: Creutzfeldt-Jakob disease associated with cadaveric dura mater grafts–Japan, January 1979–May 1996. *MMWR* 46:1066–1069, 1997.

259. Kimberlin R.H.: Reflections on the nature of the scrapie agent. *Trends Biochem. Sci.* 7:392–394, 1982.

260. Dickinson A.G. and Outram G.W.: Genetic aspects of unconventional virus infections: the basis of the virino hypothesis. *Novel Infectious Agents and the Central Nervous System. Ciba Foundation Symposium 135.* Bock G. and Marsh J., Eds. John Wiley and Sons, Chichester, UK, 1988 pp. 63–83.

261. Bruce M.E., McConnell I., Fraser H., and Dickinson A.G.: The disease characteristics of different strains of scrapie in *Sinc* congenic mouse lines: implications for the nature of the agent and host control of pathogenesis. *J. Gen. Virol.* 72:595–603, 1991.

262. Weissmann C.: A "unified theory" of prion propagation. *Nature* 352:679–683, 1991.

263. Ridley R.M. and Baker H.F.: To what extent is strain variation evidence for an independent genome in the agent of the transmissible spongiform encephalopathies? *Neurodegeneration* 5:219–231, 1996.

264. Fraser H. and Dickinson A.G.: Scrapie in mice. Agent-strain differences in the distribution and intensity of grey matter vacuolation. *J. Comp. Pathol.* 83:29–40, 1973.

265. Bruce M.E., McBride P.A., and Farquhar C.F.: Precise targeting of the pathology of the sialoglycoprotein, PrP, and vacuolar degeneration in mouse scrapie. *Neurosci. Lett.* 102:1–6, 1989.

266. DeArmond S.J., Yang S.-L., Lee A., Bowler R., Taraboulos A., Groth D., and Prusiner S.B.: Three scrapie prion isolates exhibit different accumulation patterns of the prion protein scrapie isoform. *Proc. Natl. Acad. Sci. USA* 90:6449–6453, 1993.

267. Dickinson A.G. and Meikle V.M.: A comparison of some biological characteristics of the mouse-passaged scrapie agents, 22A and ME7. *Genet. Res.* 13:213–225, 1969.

268. Bruce M.E. and Dickinson A.G.: Biological evidence that the scrapie agent has an independent genome. *J. Gen. Virol.* 68:79–89, 1987.

269. Hunter N., Hope J., McConnell I., and Dickinson A.G.: Linkage of the scrapie-associated fibril protein (PrP) gene and Sinc using congenic mice and restriction fragment length polymorphism analysis. *J. Gen. Virol.* 68:2711–2716, 1987.

270. Bessen R.A. and Marsh R.F.: Identification of two biologically distinct strains of transmissible mink encephalopathy in hamsters. *J. Gen. Virol.* 73:329–334, 1992.

271. Bessen R.A., Kocisko D.A., Raymond G.J., Nandan S., Lansbury P.T., and Caughey B.: Non-genetic propagation of strain-specific properties of scrapie prion protein. *Nature* 375:698–700, 1995.

272. Monari L., Chen S.G., Brown P., Parchi P., Petersen R.B., Mikol J., Gray F., Cortelli P., Montagna P., Ghetti B., Goldfarb L.G., Gajdusek D.C., Lugaresi E., Gambetti P., and Autilio-Gambetti L.: Fatal familial insomnia and familial Creutzfeldt-Jakob disease: different prion proteins determined by a DNA polymorphism. *Proc. Natl. Acad. Sci. USA* 91:2839–2842, 1994.

273. Parchi P., Castellani R., Capellari S., Ghetti B., Young K., Chen S.G., Farlow M., Dickson D.W., Sima A.A.F., Trojanowski J.Q., Petersen R.B., and Gambetti P.: Molecular basis of phenotypic variability in sporadic Creutzfeldt-Jakob disease. *Ann. Neurol.* 39:767–778, 1996.

274. Mastrianni J., Nixon F., Layzer R., DeArmond S.J., and Prusiner S.B.: Fatal sporadic insomnia: fatal familial insomnia phenotype without a mutation of the prion protein gene. *Neurology* 48 [Suppl.]:A296, 1997.

275. Fraser H. and Dickinson A.G.: The sequential development of the brain lesions of scrapie in three strains of mice. *J. Comp. Pathol.* 78:301–311, 1968.

276. Carp R.I., Meeker H., and Sersen E.: Scrapie strains retain their distinctive characteristics following passages of homogenates from different brain regions and spleen. *J. Gen. Virol.* 78:283–290, 1997.

277. DeArmond S.J., Sánchez H., Yehiely F., Qiu Y., Ninchak-Casey A., Daggett V., Camerino A.P., Cayetano J., Rogers M., Groth D., Torchia M., Tremblay P., Scott M.R., Cohen F.E., and Prusiner S.B.: Selective neuronal targeting in prion disease. *Neuron* 19:1337–1348, 1997.

278. Medori R., Tritschler H.-J., LeBlanc A., Villare F., Manetto V., Chen H.Y., Xue R., Leal S., Montagna P., Cortelli P., Tinuper P., Avoni P., Mochi M., Baruzzi A., Hauw J.J., Ott J., Lugaresi E., Autilio-Gambetti L., and Gambetti P.: Fatal familial insomnia, a prion disease with a mutation at codon 178 of the prion protein gene. *N. Engl. J. Med.* 326:444–449, 1992.

279. Anderson R.M., Donnelly C.A., Ferguson N.M., Woolhouse M.E.J., Watt C.J., Udy H.J., MaWhinney S., Dunstan S.P., Southwood T.R.E., Wilesmith J.W., Ryan J.B.M., Hoinville L.J., Hillerton J.E., Austin A.R., and Wells G.A.H.: Transmission dynamics and epidemiology of BSE in British cattle. *Nature* 382:779–788, 1996.

280. Nathanson N., Wilesmith J., and Griot C.: Bovine spongiform encephalopathy (BSE): cause and consequences of a common source epidemic. *Am. J. Epidemiol.* 145:959–969, 1997.

281. Stekel D.J., Nowak M.A., and Southwood T.R.E.: Prediction of future BSE spread. *Nature* 381:119, 1996.

282. Wilesmith J.W., Ryan J.B.M., and Atkinson M.J.: Bovine spongiform encephalopathy–Epidemiologic studies on the origin. *Vet. Rec.* 128:199–203, 1991.

283. Wilesmith J.W.: The epidemiology of bovine spongiform encephalopathy. *Semin. Virol.* 2:239–245, 1991.

284. Kimberlin R.H.: Speculations on the origin of BSE and the epidemiology of CJD. *Bovine Spongiform Encephalopathy: The BSE Dilemma*. Gibbs C.J., Jr., Ed. Springer, New York, 1996 pp. 155–175.

285. Goldmann W., Hunter N., Manson J., and Hope J.: The PrP gene of the sheep, a natural host of scrapie. *Proceedings of the VIIIth International Congress of Virology, Berlin, Aug. 26–31* 284, 1990.

286. Goldmann W., Hunter N., Martin T., Dawson M., and Hope J.: Different forms of the bovine PrP gene have five or six copies of a short, G-C-rich element within the protein-coding exon. *J. Gen. Virol.* 72:201–204, 1991.

287. Prusiner S.B., Fuzi M., Scott M., Serban D., Serban H., Taraboulos A., Gabriel J.-M., Wells G., Wilesmith J., Bradley R., DeArmond S.J., and Kristensson K.: Immunologic and molecular biological studies of prion proteins in bovine spongiform encephalopathy. *J. Infect. Dis.* 167:602–613, 1993.

288. Hunter N., Goldmann W., Smith G., and Hope J.: Frequencies of PrP gene variants in healthy cattle and cattle with BSE in Scotland. *Vet. Rec.* 135:400–403, 1994.

289. Fraser H., McConnell I., Wells G.A.H., and Dawson M.: Transmission of bovine spongiform encephalopathy to mice. *Vet. Rec.* 123:472, 1988.

290. Dawson M., Wells G.A.H., and Parker B.N.J.: Preliminary evidence of the experimental transmissibility of bovine spongiform encephalopathy to cattle. *Vet. Rec.* 126:112–113, 1990.

291. Dawson M., Wells G.A.H., Parker B.N.J., and Scott A.C.: Primary parenteral transmission of bovine spongiform encephalopathy to the pig. *Vet. Rec.* 127:338, 1990.

292. Bruce M., Chree A., McConnell I., Foster J., and Fraser H.: Transmissions of BSE, scrapie and related diseases to mice (Abstr.). *Proceedings of the IXth International Congress of Virology* 93, 1993.

293. Bruce M.E., Will R.G., Ironside J.W., McConnell I., Drummond D., Suttie A., McCardle L., Chree A., Hope J., Birkett C., Cousens S., Fraser H., and Bostock C.J.: Transmissions to mice indicate that 'new variant' CJD is caused by the BSE agent. *Nature* 389:498–501, 1997.

294. Robinson M.M., Hadlow W.J., Knowles D.P., Huff T.P., Lacy P.A., Marsh R.F., and

Gorham J.R.: Experimental infection of cattle with the agents of transmissible mink encephalopathy and scrapie. *J. Comp. Path.* 113:241–251, 1995.

295. Hope J., Reekie L.J.D., Hunter N., Multhaup G., Beyreuther K., White H., Scott A.C., Stack M.J., Dawson M., and Wells G.A.H.: Fibrils from brains of cows with new cattle disease contain scrapie-associated protein. *Nature* 336:390–392, 1988.

296. Serban D., Taraboulos A., DeArmond S.J., and Prusiner S.B.: Rapid detection of Creutzfeldt-Jakob disease and scrapie prion proteins. *Neurology* 40:110–117, 1990.

297. Grathwohl K.-U.D., Horiuchi M., Ishiguro N., and Shinagawa M.: Sensitive enzyme-linked immunosorbent assay for detection of PrP^{Sc} in crude tissue extracts from scrapie-affected mice. *J. Virol. Methods* 64:205–216, 1997.

298. Scott M.R., Safar J., Telling G., Nguyen O., Groth D., Torchia M., Koehler R., Tremblay P., Walther D., Cohen F.E., DeArmond S.J., and Prusiner S.B.: Identification of a prion protein epitope modulating transmission of bovine spongiform encephalopathy prions to transgenic mice. *Proc. Natl. Acad. Sci. USA* 94:14279–14284, 1997.

299. Taylor K.C.: The control of bovine spongiform encephalopathy in Great Britain. *Vet. Rec.* 129:522–526, 1991.

300. Fraser H., Bruce M.E., Chree A., McConnell I., and Wells G.A.H.: Transmission of bovine spongiform encephalopathy and scrapie to mice. *J. Gen. Virol.* 73:1891–1897, 1992.

301. Hunter G.D., Millson G.C., and Chandler R.L.: Observations on the comparative infectivity of cellular fractions derived from homogenates of mouse-scrapie brain. *Res. Vet. Sci.* 4:543–549, 1963.

302. Eklund C.M., Kennedy R.C., and Hadlow W.J.: Pathogenesis of scrapie virus infection in the mouse. *J. Infect. Dis.* 117:15–22, 1967.

303. Will R.G., Ironside J.W., Zeidler M., Cousens S.N., Estibeiro K., Alperovitch A., Poser S., Pocchiari M., Hofman A., and Smith P.G.: A new variant of Creutzfeldt-Jakob disease in the UK. *Lancet* 347:921–925, 1996.

304. Bateman D., Hilton D., Love S., Zeidler M., Beck J., and Collinge J.: Sporadic Creutzfeldt-Jakob disease in a 18-year-old in the UK (Lett.). *Lancet* 346:1155–1156, 1995.

305. Britton T.C., Al-Sarraj S., Shaw C., Campbell T., and Collinge J.: Sporadic Creutzfeldt-Jakob disease in a 16-year-old in the UK (Lett.). *Lancet* 346:1155, 1995.

306. Ironside J.W.: The new variant form of Creutzfeldt-Jakob disease: a novel prion protein amyloid disorder (Editorial). *Amyloid: Int. J. Exp. Clin. Invest.* 4:66–69, 1997.

307. Chazot G., Broussolle E., Lapras C.I., Blättler T., Aguzzi A., and Kopp N.: New variant of Creutzfeldt-Jakob disease in a 26-year-old French man. *Lancet* 347:1181, 1996.

308. CDC: Surveillance for Creutzfeldt-Jakob Disease–United States. *MMWR* 45:665–668, 1996.

309. Baker H.F., Ridley R.M., and Wells G.A.H.: Experimental transmission of BSE and scrapie to the common marmoset. *Vet. Rec.* 132:403–406, 1993.

310. Lasmézas C.I., Deslys J.-P., Demaimay R., Adjou K.T., Lamoury F., Dormont D., Robain O., Ironside J., and Hauw J.-J.: BSE transmission to macaques. *Nature* 381:743–744, 1996.

311. Collinge J., Palmer M.S., Sidle K.C., Hill A.F., Gowland I., Meads J., Asante E., Bradley R., Doey L.J., and Lantos P.L.: Unaltered susceptibility to BSE in transgenic mice expressing human prion protein. *Nature* 378:779–783, 1995.

312. Cousens S.N., Vynnycky E., Zeidler M., Will R.G., and Smith P.G.: Predicting the CJD epidemic in humans. *Nature* 385:197–198, 1997.

313. Raymond G.J., Hope J., Kocisko D.A., Priola S.A., Raymond L.D., Bossers A., Ironside J., Will R.G., Chen S.G., Petersen R.B., Gambetti P., Rubenstein R., Smits M.A., Lansbury P.T., Jr., and Caughey B.: Molecular assessment of the potential transmissibilities of BSE and scrapie to humans. *Nature* 388:285–288, 1997.

314. Collinge J., Sidle K.C.L., Meads J., Ironside J., and Hill A.F.: Molecular analysis of prion strain variation and the aetiology of "new variant" CJD. *Nature* 383:685–690, 1996.

315. Hill A.F., Desbruslais M., Joiner S., Sidle K.C.L., Gowland I., Collinge J., Doey L.J., and Lantos P.: The same prion strain causes vCJD and BSE.. *Nature* 389:448–450, 1997.

316. Parchi P., Capellari S., Chen S.G., Petersen R.B., Gambetti P., Kopp P., Brown P., Kitamoto T., Tateishi J., Giese A., and Kretzschmar H.: Typing prion isoforms (Lett.). *Nature* 386:232–233, 1997.

317. Somerville R.A., Chong A., Mulqueen O.U., Birkett C.R., Wood S.C.E.R., and Hope J.: Biochemical typing of scrapie strains. *Nature* 386:564, 1997.

318. Haraguchi T., Fisher S., Olofsson S., Endo T., Groth D., Tarantino A., Borchelt D.R., Teplow D., Hood L., Burlingame A., Lycke E., Kobata A., and Prusiner S.B.: Asparagine-linked glycosylation of the scrapie and cellular prion proteins. *Arch. Biochem. Biophys.* 274:1–13, 1989.

319. Endo T., Groth D., Prusiner S.B., and Kobata A.: Diversity of oligosaccharide structures linked to asparagines of the scrapie prion protein. *Biochemistry* 28:8380–8388, 1989.

320. Wickner R.B.: [URE3] as an altered URE2 protein: evidence for a prion analog in *Saccharomyces cerevisiae. Science* 264:566–569, 1994.

321. Chernoff Y.O., Lindquist S.L., Ono B., Inge-Vechtomov S.G., and Liebman S.W.: Role of the chaperone protein Hsp104 in propagation of the yeast prion-like factor [*psi*⁺]. *Science* 268:880–884, 1995.

322. Glover J.R., Kowal A.S., Schirmer E.C., Patino M.M., Liu J.-J., and Lindquist S.: Self-seeded fibers formed by Sup35, the protein determinant of [*PSI*⁺], a heritable prion-like factor of S. cerevisiae. *Cell* 89:811–819, 1997.

323. King C.-Y., Tittman P., Gross H., Gebert R., Aebi M., and Wüthrich K.: Prion-inducing domain 2–114 of yeast Sup35 protein transforms *in vitro* into amyloid-like filaments. *Proc. Natl. Acad. Sci. USA* 94:6618–6622, 1997.

324. Paushkin S.V., Kushnirov V.V., Smirnov V.N., and Ter-Avanesyan M.D.: In vitro propagation of the prion-like state of yeast Sup35 protein. *Science* 277:381–383, 1997.

325. Coustou V., Deleu C., Saupe S., and Begueret J.: The protein product of the *het-s* heterokaryon incompatibility gene of the fungus *Podospora anserina* behaves as a prion analog. *Proc. Natl. Acad. Sci. USA* 94:9773–9778, 1997.

326. Derkatch I.L., Chernoff Y.O., Kushnirov V.V., Inge-Vechtomov S.G., and Liebman S.W.: Genesis and variability of [*PSI*] prion factors in *Saccharomyces cerevisiae. Genetics* 144:1375–1386, 1996.

327. Patino M.M., Liu J.-J., Glover J.R., and Lindquist S.: Support for the prion hypothesis for inheritance of a phenotypic trait in yeast. *Science* 273:622–626, 1996.

328. Wickner R.B.: A new prion controls fungal cell fusion incompatibility [Commentary]. *Proc. Natl. Acad. Sci. USA* 94:10012–10014, 1997.

329. Sonneborn T.M.: The determination of hereditary antigenic differences in genically identical Paramecium cells. *Proc. Natl. Acad. Sci. USA* 34:413–418, 1948.

330. Landman O.E.: The inheritance of acquired characteristics. *Annu. Rev. Genetics* 25:1–20, 1991.

331. Beisson J. and Sonneborn T.M.: Cytoplasmid inheritance of the organization of the cell cortex of *Paramecium aurelia. Proc. Natl. Acad. Sci. USA* 53:275–282, 1965.

332. Frankel J.: Positional order and cellular handedness. *J. Cell Sci.* 97:205–211, 1990.

333. Chapman J., Ben-Israel J., Goldhammer Y., and Korczyn A.D.: The risk of developing Creutzfeldt-Jakob disease in subjects with the *PRNP* gene codon 200 point mutation. *Neurology* 44:1683–1686, 1994.

334. Spudich S., Mastrianni J.A., Wrensch M., Gabizon R., Meiner Z., Kahana I., Rosenmann H., Kahana E., and Prusiner S.B.: Complete penetrance of Creutzfeldt-Jakob disease in Libyan Jews carrying the E200K mutation in the prion protein gene. *Mol. Med.* 1:607–613, 1995.

335. Aguzzi A. and Weissmann C.: Prion research: the next frontiers. *Nature* 389:795–798, 1997.

336. Gorodinsky A. and Harris D.A.: Glycolipid-anchored proteins in neuroblastoma cells form detergent-resistant complexes without caveolin. *J. Cell Biol.* 129:619–627, 1995.

337. Goldmann W., Hunter N., Smith G., Foster J., and Hope J.: PrP genotype and agent effects in scrapie: change in allelic interaction with different isolates of agent in sheep, a natural host of scrapie. *J. Gen. Virol.* 75:989–995, 1994.

338. Belt P.B.G.M., Muileman I.H., Schreuder B.E.C., Ruijter J.B., Gielkens A.L.J., and Smits M.A.: Identification of five allelic variants of the sheep PrP gene and their association with natural scrapie. *J. Gen. Virol.* 76:509–517, 1995.

339. Clousard C., Beaudry P., Elsen J.M., Milan D., Dussaucy M., Bounneau C., Schelcher F., Chatelain J., Launay J.M., and Laplanche J.L.: Different allelic effects of the codons 136 and 171 of the prion protein gene in sheep with natural scrapie. *J. Gen. Virol.* 76:2097–2101, 1995.

340. Ikeda T., Horiuchi M., Ishiguro N., Muramatsu Y., Kai-Uwe G.D., and Shinagawa M.: Amino acid polymorphisms of PrP with reference to onset of scrapie in Suffolk and Corriedale sheep in Japan. *J. Gen. Virol.* 76:2577–2581, 1995.

341. Hunter N., Moore L., Hosie B.D., Dingwall W.S., and Greig A.: Association between natural scrapie and PrP genotype in a flock of Suffolk sheep in Scotland. *Vet. Rec.* 140:59–63, 1997.

342. Hunter N., Cairns D., Foster J.D., Smith G., Goldmann W., and Donnelly K.: Is scrapie solely a genetic disease? *Nature* 386:137, 1997.

343. O'Rourke K.I., Holyoak G.R., Clark W.W., Mickelson J.R., Wang S., Melco R.P., Besser T.E., and Foote W.C.: PrP genotypes and experimental scrapie in orally inoculated Suffolk sheep in the United States. *J. Gen. Virol.* 78:975–978, 1997.

344. Parry H.B.: *Scrapie Disease in Sheep,* Academic Press, New York, 1983.

345. Chesebro B.: Prion diseases: BSE and prions: uncertainties about the agent. *Science* 279:42–43, 1998.

346. Shinde U.P., Liu J.J., and Inouye M.: Protein memory through altered folding mediated by intramolecular chaperones. *Nature* 389:520-522, 1997.